

Searching for the Autoimmune Thyroid Disease Susceptibility Genes: From Gene Mapping to Gene Function

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The autoimmune thyroid diseases (AITD) are complex diseases that are caused by an interaction between susceptibility genes and environmental triggers. Genetic susceptibility, in combination with external factors (e.g., dietary iodine), is believed to initiate the autoimmune response to thyroid antigens. Abundant epidemiological data, including family and twin studies, point to a strong genetic influence on the development of AITD. Various techniques have been used to identify the genes contributing to the etiology of AITD, including candidate gene analysis and whole genome screening. These studies have enabled the identification of several loci (genetic regions) that are linked with AITD, and in some of these loci putative AITD susceptibility genes have been identified. Some

of these genes/loci are unique to Graves' disease (GD) and Hashimoto's thyroiditis (HT), and some are common to both diseases, indicating that there is a shared genetic susceptibility to GD and HT. The putative GD and HT susceptibility genes include both immune modifying genes (e.g., human leukocyte antigen, cytotoxic T lymphocyte antigen-4) and thyroid-specific genes (e.g., TSH receptor, thyroglobulin). Most likely these loci interact, and their interactions may influence disease phenotype and severity. It is hoped that in the near future additional AITD susceptibility genes will be identified and the mechanisms by which they induce AITD will be unraveled. (*Endocrine Reviews* 24: 694–717, 2003)

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I. Introduction

THE AUTOIMMUNE THYROID diseases (AITD) encompass a number of conditions that have in common cellular and humoral immune responses targeted at the thyroid gland. The classical AITD include Graves' disease (GD) and Hashimoto's thyroiditis (HT), both of which involve infiltration of the thyroid by T and B cells reactive with thyroid antigens, and production of thyroid autoantibodies, with the resultant clinical manifestations (hyperthyroidism in GD and hypothyroidism in HT) (reviewed in Refs. 1 and 2). Additional variants of AITD include postpartum thyroiditis (reviewed in Refs. 3–5), drug-induced thyroiditis [e.g.,

Abbreviations: AITD, Autoimmune thyroid disease(s); APC, antigen-presenting cell(s); CTLA-4, cytotoxic T lymphocyte antigen-4; DM, diabetes mellitus; DZ, dizygotic; EAT, experimental autoimmune thyroiditis; GD, Graves' disease; GO, Graves' ophthalmopathy; HLA, human leukocyte antigen; HLOD, heterogeneity LOD score; HT, Hashimoto's thyroiditis; IDDM, insulin-dependent IDDM; IgH, IgG heavy chain; LD, linkage disequilibrium; LOD, logarithm of odds; MHC, major histocompatibility complex; MLS, maximum LOD score; MZ, monozygotic; NHANES III, National Health and Nutrition Examination Survey; RR, relative risk; SNP, single nucleotide polymorphism; TAb, thyroid antibody or antibodies; TDT, transmission disequilibrium test; Tg, thyroglobulin; Tg-Ab, Tg antibodies; Tgms2, Tg microsatellites in intron 27; TPO, thyroid peroxidase; TPO-Ab, TPO antibodies; TSHR, TSH receptor; UTR, untranslated region.

Amiodarone (for review, see Ref. 6), and interferon- α (7, 8)], thyroiditis accompanying the polyglandular autoimmune syndromes (reviewed in Refs. 9 and 10), and the presence of thyroid antibodies (TAb) [please note that we use the abbreviation TAb for thyroid peroxidase (TPO) and thyroglobulin (Tg) antibodies only] with no apparent clinical disease (11) (in this review we use the term AITD to denote GD and HT only). Although the exact etiology of the immune response to the thyroid remains unknown, there is solid evidence for a major genetic influence on the development of AITD (reviewed in Refs. 12 and 13). Therefore, the current paradigm is that AITD are complex diseases in which susceptibility genes and environmental triggers act in concert to initiate the autoimmune response to the thyroid. In this review, we will summarize the recent advances in our understanding of the genetic contribution to the development of AITD. We will limit the discussion to the genetics of GD, HT, and thyroid autoantibodies.

II. The Autoimmune Thyroid Diseases Are Familial

A. Population data (Table 1)

A recently published, large, epidemiological study in the United States population, which is iodine sufficient [National Health and Nutrition Examination Survey (NHANES III)], found the prevalence of thyrotoxicosis from any cause to be 1.3% (0.5% clinical and 0.7% subclinical) (14). Another recent large study from the United States (The Colorado Thyroid Disease Prevalence Study) found the prevalence of subclinical and clinical hyperthyroidism to be 2.1 and 0.1%, respectively (15). Additionally, a comprehensive survey in the town of Whickham in the United Kingdom found the prevalence of thyrotoxicosis to be 2.7% in women, and 10-fold less in men (0.16–0.23%) (16). Because the commonest cause of thyrotoxicosis in iodine-sufficient Western countries is GD, ac-

counting for 70–80% of cases (17–19), these frequencies are probably good estimates of the prevalence of GD in the United Kingdom and United States. However, the level of dietary iodine has been shown to markedly affect the incidence of thyrotoxicosis and GD (20). In iodine-deficient regions, thyrotoxicosis is more prevalent, and GD accounts for a lower percentage of all thyrotoxicosis cases. Indeed, a recent well-designed large study from Denmark has shown the incidence of thyrotoxicosis to be 65.4/100,000/yr in a mild iodine deficiency region, and 92.9/100,000/yr in an area with moderate iodine deficiency (21). In another study, the incidence of GD was higher (19.7/100,000/yr) in Iceland (a high iodine intake region) when compared with a region in Denmark with low average iodine intake (14.8/100,000/yr) (20).

Epidemiological surveys from various, mostly iodine-sufficient regions have shown a relatively similar incidence of GD in Caucasian populations (18, 19, 22–24). The annual incidence of GD in these studies was approximately 20–25 per 100,000 (Table 1).

In mostly iodine-sufficient regions, similar prevalence and incidence trends have also been observed for HT. In the NHANES III study, the prevalence of hypothyroidism was found to be 4.6% (0.3% clinical and 4.3% subclinical) (14). In the Whickham survey, the prevalence of spontaneous hypothyroidism was 1.5% in females and less than 0.1% in males (16). These prevalence rates are similar to those reported in Finland (25), Japan (26), and in another U.S. survey (27). A higher prevalence of hypothyroidism was reported in the Colorado study (0.4% clinical and 9.0% subclinical), but this study included an older population with a higher percentage of women than in the general population of Colorado (15), and this might have increased the prevalence of subclinical hypothyroidism in this study (28). It should be emphasized that the phenotype definitions of GD and HT in these epidemiological studies are not always specified and

TABLE 1. Prevalence and incidence of clinical thyrotoxicosis/GD and hypothyroidism/HT in different geographic regions

Country	Phenotype	Years of survey	Prevalence (F/M)	Incidence per 100,000/yr (F/M)	Ref.
Sweden	GD	1970–1974	NA	17.7 (27.2/7.4)	19
Sweden	GD	1988–1990	NA	22.2 (34.4/8.8)	30
United States (Mayo)	GD	1935–1967	NA	(36.8/8.3)	29
United States (Mayo)	HT	1935–1967	NA	(40.7/1.0)	29
Iceland	Thyrotoxicosis	1980–1982	NA	23.6 (38.4/8.9)	22
New Zealand	Thyrotoxicosis	1983–1985	NA	25.8 (40.7/10.5)	18
United Kingdom (Whickham 1)	Thyrotoxicosis	1972–1974	(2.7%/0.16%)	NA	16
United Kingdom (Whickham 2)	Thyrotoxicosis	1972–1992	(3.9%/0.2%)	(80/0) ^a	159
United States (NHANES III)	Thyrotoxicosis	1988–1994	0.5% ^b	NA	14
United States (Colorado)	Thyrotoxicosis	1995	0.1% ^b	NA	15
United States (Oakland)	Thyrotoxicosis	1977	0.31% ^b	NA	289
United States ^c	Thyrotoxicosis	1965–1995	1.2%	13.9	27
United Kingdom (Whickham 1)	Hypothyroidism	1972–1974	(1.9%/<0.1%)	NA	16
United Kingdom (Whickham 2)	Hypothyroidism	1972–1992	(7.7%/1.3%)	(350/60) ^a	159
United States (NHANES III)	Hypothyroidism	1988–1994	0.3% ^b	NA	14
United States (Colorado)	Hypothyroidism	1995	0.4% ^b	NA	15
United States (Oakland)	Hypothyroidism	1977	0.50% ^b	NA	289
United States ^c	Hypothyroidism	1965–1995	0.8%	21.8	27

The incidence/prevalence of subclinical disease is not included in the table. See text (Section II.A) for details. F, female; M, male; NA, not available.

^a The incidence was calculated only in the survivors of the original study and may not reflect the incidence in the general population.

^b Clinical hypo/hyperthyroidism.

^c This is a metaanalysis of several studies.

that variations in phenotype definitions may significantly affect the population data.

The comparable prevalence and incidence of GD and HT in geographically different populations that are iodine sufficient suggests a significant genetic effect on the development of these diseases because these populations are exposed to different environmental factors (other than dietary iodine). Moreover, it may imply that different Caucasian populations share some of their susceptibility genes for AITD.

Longitudinal surveys can also point to the relative importance of genetic factors in the etiology of a disease. An early longitudinal study from the Mayo Clinic (1935–1967) showed no significant change in the incidence of GD over the 33 yr of the study (29). The stable incidence of GD with time points to a genetic susceptibility to GD because the genetic makeup of a population does not change over several decades, but environmental factors would be expected to vary over time. Therefore, the incidence rates of diseases with strong environmental influences (*e.g.*, lung cancer) are expected to change over time, whereas the incidence rates of diseases with strong genetic influences should be more stable with time, although complex diseases in which environmental and genetic factors interact can also vary with time. The Mayo Clinic observations were supported by a more recent study from Sweden (30) that showed no significant change in the incidence of GD in Malmo over a 20-yr period (the incidence was 17.7/100,000/yr in 1970–1974 and 22.2/100,000/yr in 1988–1990). However, the Swedish study also found an increased incidence of GD in females younger than 50 yr (30), demonstrating that environmental effects also play a role in the etiology of GD. In contrast to GD, the early Mayo survey (1935–1967) found a significant increase in the incidence of HT over the 33 yr of the survey (29). This may have reflected a stronger environmental influence on the development of HT or, more likely, a change in the diagnostic criteria over time. Indeed, the diagnosis of HT is much more subject to variation (*e.g.*, goitrous *vs.* atrophic) than the diagnosis of GD (31).

B. Family studies

The familial occurrence of AITD has been reported by investigators for many years. Earlier studies showing familial aggregation of AITD were mostly observational, based on careful family histories from patients. These early studies reported a family history of thyroid disease in up to 60% of patients with GD (32, 33). Later, in the 1960s, Hall and Stanbury (34) showed that 33% of siblings of patients with GD or HT developed AITD themselves. Additionally, they found that 56% of siblings of AITD patients had TAb, and in almost all cases at least one of the parents of an affected individual had TAb, suggesting dominant inheritance of the TAb trait (34). More recently, several groups have reported a high frequency of thyroid abnormalities in relatives of patients with GD (35–38), most commonly the presence of thyroid autoantibodies that were reported in up to 50% of the siblings of patients with GD (reviewed in Ref. 39; also see Refs. 36 and 40). A recent survey by our own group revealed that 41 of 114 (36%) GD patients with ophthalmopathy reported a family

history of AITD, and 26 of 114 (23%) had a first-degree relative with AITD (41).

C. Sibling risk ratio (λ_s)

Familial clustering of a disease does not necessarily mean that the disease is genetic. Familial clustering can be the result of random chance, shared extrinsic (environmental) factors, shared intrinsic (genetic) factors, or a combination of these. Coincidental familial clustering may occur in cases in which a disease is common and, therefore, may occur in multiple members of the same family by chance alone. For example, essential hypertension is common in Western populations and, therefore, may occur in multiple members in the same family. However, this does not necessarily mean that it is caused by genetic factors. Shared external factors include diet (*e.g.*, iodine deficiency can cause familial clustering of cretinism and goiter), infections [*e.g.*, exposure to certain viruses can cause familial clustering of subacute thyroiditis (reviewed in Ref. 42)], and other environmental exposures [*e.g.*, exposure to radiation fallout can cause familial clustering of childhood thyroid cancer (43)]. These situations have to be distinguished from genetic causes of familial clustering.

Several methods are available to determine whether familial clustering of a disease is the result of genetic susceptibility or nongenetic factors. One method is to calculate the λ_s , which is the ratio of the prevalence of the disease in siblings of affected individuals compared with the prevalence of the disease in the general population (44). The λ_s expresses the increased risk of developing the disease in an individual who has a sibling with the disease and is a quantitative measure of the genetic contribution to the disease. A λ_s greater than 5 usually indicates a significant genetic contribution to the pathogenesis of a disease (44). In the case of the AITD, the λ_s was estimated to be between 5.9 and more than 10 (12, 41, 45, 46), supporting a strong genetic influence on the development of AITD.

D. Segregation analyses

Another approach for determining whether familial clustering of a disease reflects genetic predisposition is to perform segregation analyses. Here, the segregation of the disease in families is analyzed to see whether it occurs at random or demonstrates Mendelian or a complex pattern of inheritance. This analysis has the advantage of determining both the existence of genetic predisposition to a disease and the mode of inheritance. Two segregation analyses have been performed in families with TAb and have suggested a Mendelian dominant pattern of inheritance for the tendency to develop TAb (47–49). In keeping with these observations, it was recently found that recognition of particular TPO epitopes within the autoantibody immunodominant region may be transmitted within families (50).

E. Twin studies (Table 2)

The most powerful method for evaluating genetic predisposition to complex diseases involves twins. Twin studies are based on comparison of the concordance (simultaneous oc-

TABLE 2. Results of some of the recent twin studies in AITD

Phenotype	MZ twins		DZ twins		Ref.
	No. of pairs	Concordant (%)	No. of pairs	Concordant (%)	
GD	18	4 (22)	33	0 (0)	53
GD	37	8 (22)	59	1 (1.7)	54
GD	35	6 (17)	54	1 (1.9)	55
HT	8	3 (38)	13	0 (0)	56
GD/HT	35	11 (31)	54	2 (3.7)	55
Tg-Ab	17	10 (59)	13	3 (23)	57
TPO-Ab	15	7 (47)	14	4 (29)	57
TAb	5	4 (80)	10	4 (40)	56

currence) of a given disease among monozygotic (MZ; *i.e.*, near identical) twins with the concordance among dizygotic (DZ; *i.e.*, fraternal) twins. MZ twins have almost identical genetic makeup, whereas DZ twins share 50% of their genes (like siblings). Therefore, if the concordance is higher in the MZ twins when compared with the DZ twins, it suggests that the disease has an inherited component, although the definition of identical twins may not be as simple as first thought (for a review, see Ref. 51). Any discordance among the MZ twin pairs is usually interpreted to mean that the gene or genes concerned show reduced penetrance, *i.e.*, certain non-genetic events must occur or certain environmental factors must be present before the disease becomes manifest. It must be emphasized that MZ twins are not identical in their immune repertoire due to somatic recombinations that T and B cells undergo throughout life, as well as individual immune experiences that influence the immune repertoire (52). Therefore, part of the observed discordance between MZ twins may also be due to the discordance in their immune repertoire. Several large twin studies have been reported from Denmark showing a higher concordance of AITD in MZ twins when compared with DZ twins (Table 2). For GD, the concordance was 35% in MZ twins and 3% in DZ twins (53, 54). A recent GD twin study from California confirmed the Danish twin study results (55). Brix *et al.* (54) have suggested that genetic factors are responsible for 79% of the susceptibility to develop GD, whereas environmental factors contribute the remaining 21%, but this requires further evaluation.

Twin studies in HT have shown concordance rates of 55 and 0% in MZ and DZ twins, respectively (56). The concordance rates for thyroid autoantibodies (TAb) were also reported to be higher in MZ twins compared with DZ twins (Table 2). In a recent study from the United Kingdom, the concordance rates for Tg antibodies (Tg-Ab) were 59 and 23% for MZ and DZ twins, respectively (57). The concordance rates for TPO antibodies (TPO-Ab) were 47 and 29% for MZ and DZ twins, respectively (57). The Danish twin studies also found higher concordance rates for TAb in MZ twins (80%) when compared with DZ twins (40%) (56). These twin data confirm with remarkable clarity the presence of a substantial inherited susceptibility to AITD.

III. Tools Used to Map and Identify Complex Disease Genes

Based on the abundant epidemiological evidence for a strong genetic effect on the development of AITD, several

groups have been trying to map and identify the AITD susceptibility genes. The two basic strategies used for mapping complex disease genes include linkage and association studies of candidate genes and whole genome screening. These tools have been successful in mapping classical Mendelian disorders [*e.g.*, Pendred's syndrome (58)], and more recently have enabled the identification of the first complex disease genes for type 1 diabetes (59) and Crohn's disease (60). These strategies have also been used in the study of the genetic susceptibility to AITD.

A. Linkage and association

1. *Linkage.* Genetic linkage techniques are powerful tools for analyzing complex disease-related genes because they detect genes that have a major influence on the development of a disease (61). However, linkage studies are less sensitive than association studies because they do not detect less influential genes (61). A linkage study, therefore, may be negative in the absence of major genes contributing to disease susceptibility. The principle of linkage analysis is based on the fact that if two genes or markers are close together on a chromosome, they will cosegregate because the likelihood that a recombination will occur between them during meiosis is low. Therefore, if a tested marker is close to a disease susceptibility gene, its alleles will cosegregate with the disease in families (Fig. 1). The logarithm of odds (LOD) score is the measure of the likelihood of linkage between a disease and a genetic marker (62). The LOD score is the base-10 logarithm of the odds ratio in favor of linkage. In Mendelian disorders, a LOD score of greater than 3 (*i.e.*, odds ratio greater than 1000) is considered strong evidence for linkage (62). The classical linkage tests are model based (parametric), *i.e.*, different modes of inheritance and penetrance have to be tested when calculating the likelihood of linkage. The parametric tests are the most powerful statistical tests for linkage (63, 64), and they can be used to test for heterogeneity within a dataset (heterogeneity exists in a dataset when more than one gene causes the same disease phenotype; see Section III.A.3) (65). In complex diseases, the mode of inheritance is often unknown, and therefore, simpler model-independent methods (nonparametric) have also been widely used (44). One such method is sib-pair analysis (44). In this method, siblings that are both affected by the disease being studied are tested for sharing of alleles at a marker locus. By random chance alone, the sibs would be expected to share one allele of the marker 50% of the time and two alleles 25% of the time. If affected sib-pairs share a significantly higher than expected proportion of alleles at the marker locus, this suggests that the region containing the marker locus also contains the disease gene. The observed to expected allele sharing can then be converted to a LOD score equivalent.

2. *Guideline for measuring linkage in complex diseases.* In simple Mendelian disorders, a maximum LOD score (MLS) of more than 3 is considered strong evidence of linkage (62). However, the inheritance of complex diseases (*e.g.*, AITD) does not follow a simple Mendelian pattern. These diseases are likely to be caused by several genes and have reduced penetrance (*i.e.*, not all the individuals inheriting the gene will

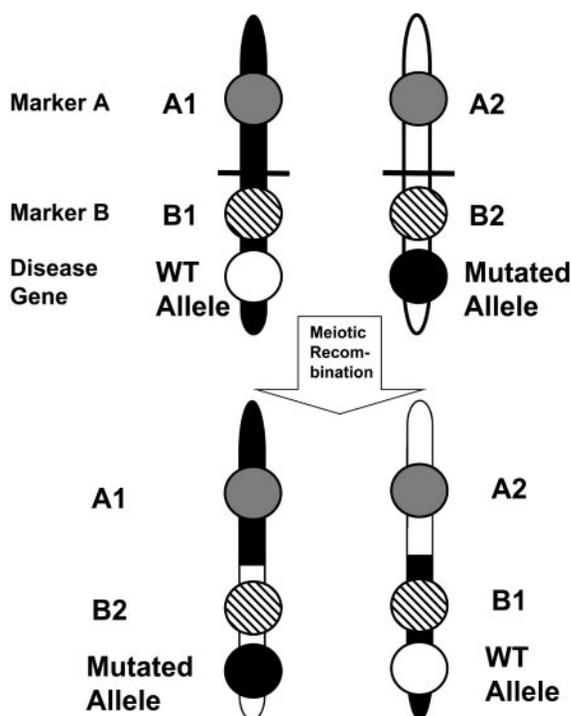


FIG. 1. The principle of linkage. During meiosis, when homologous chromosomes pair, they often break at identical points along their length and switch the distal segments to the breaking point. This results in an exchange of identical segments of the chromosome between two homologous chromosomes. The principle of linkage analysis is based on the fact that if a marker (B) is close to a susceptibility gene on a chromosome, they will segregate together during recombination, whereas a marker (A) that is far from the gene will not cosegregate with it. In the example shown, marker B is linked with the disease gene, and within a given family a certain allele of marker B (B2 in this example) will be seen more frequently than expected by random chance in the affected individuals (*i.e.*, those inheriting the mutated allele of the gene). The closer the marker is to the disease gene, the stronger the linkage with the disease [wild-type (WT) allele]. [Adapted with permission from Y. Tomer and T. F. Davies: Oxford Textbook of Endocrinology and Diabetes (edited by J. A. H. Wass and S. M. Shalet), Oxford University Press, Oxford, UK, 2002, p 358 (308). © Oxford University Press.]

develop the disease). In addition, it is likely that different genes may cause almost identical phenotypes (genetic heterogeneity). This results in non-Mendelian transmission of the disease in pedigrees and makes mapping the susceptibility genes for complex diseases difficult. Therefore, Lander and Kruglyak (66) have suggested guidelines for genetic linkage studies in complex traits. According to their guidelines, in complex diseases a LOD score of greater than 1.9 is suggestive of linkage, and a LOD score of greater than 3.3 indicates significant linkage in studies using the parametric approach. For nonparametric sib-pair studies, the cutoff LOD scores are higher (66). Linkage is confirmed if evidence for linkage is replicated in two separate datasets (66). Conversely, a LOD score lower than -2.0 has been used to exclude linkage.

3. Phenotype definitions and genetic heterogeneity. Phenotype definitions are important in genetic studies because different phenotypes are likely to be caused by different genes and analyzing them together would make identification of these

genes more difficult. Although the GD phenotype is relatively homogenous, GD can occur with or without a large goiter and/or ophthalmopathy and/or pretibial myxedema, and these should be analyzed separately. However, our segregation analysis has shown that the ophthalmopathy phenotype is most likely not determined by genetic factors (41), and therefore, it is possible that the subsetting by ophthalmopathy may not be necessary. In the case of HT, the phenotype is even more variable (*e.g.*, the presence of goiter, TPO-Ab, and the need for thyroid hormone replacement therapy) (31), and therefore, strict definitions of HT should be used in genetic studies to ensure a uniform HT phenotype. However, even when the phenotype is uniform, genetic heterogeneity can still exist. Genetic heterogeneity exists when different genotypes give rise to indistinguishable phenotypes (67). If heterogeneity exists in a dataset of families with a given phenotype (*e.g.*, AITD), the dataset may include only a subset of families that are linked with a tested marker. The existence of genetic heterogeneity in a dataset can be tested for by the Admixture Test, which calculates the likelihood that a proportion of the families in a dataset are linked to the marker (the fraction of linked families is estimated by the α -statistic) (65, 68, 69). For example, the heterogeneity LOD score (HLOD) function of the Genehunter computer program performs the Admixture Test and can be used to detect heterogeneity in a given dataset (70).

4. Association. Linkage studies are excellent for screening the whole genome. However, they have limited resolution ($\sim 2\text{--}3$ cM) because as the linked interval is narrowed all markers in the region will show linkage (61). Association studies are more sensitive than linkage studies and therefore are better for fine-mapping linked genetic regions, because the association signal [*i.e.*, the relative risk (RR)] increases as the markers get closer to the susceptibility gene. Association analysis is highly sensitive and may detect genes contributing less than 5% of the total genetic contribution to a disease (71).

Association analyses are performed by comparing the frequency of the allele studied [*e.g.*, human leukocyte antigen (HLA)-DR3] in unrelated patients and in unrelated, ethnically matched controls. This is usually performed by typing each individual patient and each control for the tested marker, but recently, methods for DNA pooling have been developed, which could simplify large-scale association studies (72). If the allele tested is associated with the disease, it will appear significantly more frequently in patients than in controls. The probability of having the disease in an individual positive for the allele compared with an individual negative for the allele is estimated by the RR (73). There are at least two possible explanations for the existence of an association between an allele and a disease: 1) the associated allele itself is the genetic variant causing an increased risk for the disease; and 2) the associated allele itself is not causing the disease but rather a gene in linkage disequilibrium (LD) with it (74). Linkage disequilibrium exists when chromosomes with the mutant allele at the disease locus carry certain marker alleles more often than expected.

This population-based association method may produce spurious associations if the patients and controls are not

accurately matched (population stratification) (75). Therefore, new association tests have been developed that are family-based and use an internal control group from within each family, thus avoiding the necessity to match patients and controls altogether. The most widely used family-based association test is the transmission disequilibrium test (TDT) (75–77). The TDT is based on comparison of parental marker alleles that are transmitted and those that are not transmitted to affected children (Fig. 2). Assuming two heterozygous parents for a certain tested marker, the four parental alleles in each family are categorized into two groups: those transmitted to a child with the disease (T alleles), and those not transmitted to an affected child (N alleles). The same allele may belong to the T group or the N group in different families. The frequency of the T alleles *vs.* the N alleles is then compared by a χ^2 test. An association between a certain allele and the disease exists if there is an excess occurrence of this allele in the T group compared with the N group. The TDT can also serve as a linkage test if there is a known LD between the tested marker and the disease.

5. Power calculations. Before performing a linkage/association study, power calculations should determine whether the proposed dataset size is sufficient to detect/reject linkage/association with the marker. The weaker the association or linkage, the larger the dataset required to detect it. Power calculations for case-control association studies are performed by simulating a dataset similar to the one being used in the study. Data for a hypothetical marker locus associated with the disease are generated using a variety of odds ratios to generate proportions of patients and controls, with and without the marker. The association analyses for that marker

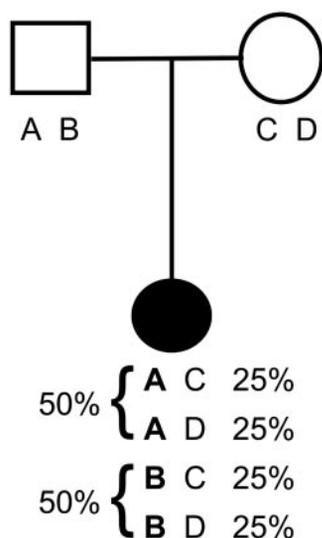


FIG. 2. TDT. In the pedigree shown, each parent is heterozygous at the tested marker. Each allele is expected to be transmitted to affected individuals from their parents 50% of the time by chance alone, and each genotype (combination of alleles) is expected to be transmitted 25% of the time. If, in a large number of pedigrees, allele A is transmitted to affected offspring more than 50% of the time (transmission distortion), then allele A is associated with the mutation in the disease gene. [Adapted with permission from Y. Tomer and T. F. Davies: Oxford Textbook of Endocrinology and Diabetes (edited by J. A. H. Wass and S. M. Shalet), Oxford University Press, Oxford, UK, 2002, p 358 (308). © Oxford University Press.]

determine the odds ratio at which an association would not be detected with the given dataset. For linkage studies, power calculations are performed by simulating a large number (>1000) of datasets consisting of families similar (or identical) to the ones being used and simulating a marker that is linked to the disease locus in these families. It is also possible to simulate another marker that is not linked to the disease in the simulated families to determine whether one can reject linkage with the given marker in the chosen families. The linkage analysis is then run on each of the datasets to find the percentage of times at which the simulated linked marker will show linkage, and similarly the percentage of times at which the simulated nonlinked marker will show evidence against linkage (78, 79). Genetic studies should always provide this analysis to show that the dataset is appropriate.

B. Candidate gene analysis

Candidate genes are of known sequence and location that by virtue of their physiological functions may be involved in disease pathogenesis. For example, one can hypothesize that the TSH receptor (TSHR) may be a candidate gene for GD because the hallmark of the disease is the presence of TSHR antibodies. If a candidate locus is indeed the cause of a disease, then markers in that locus should show association and linkage with the disease (80). Because the basic abnormality in AITD is an immune response against thyroid antigens, possible candidate genes for AITD include genes that control immune responses [*e.g.*, the major histocompatibility complex (MHC) genes, the T cell receptor genes, and antibody genes], and genes encoding the target autoantigens in AITD (Tg, TPO, iodide transporter, TSHR). Many of these genes have now been studied for a possible role in the genetic susceptibility to AITD (see Sections IV and V).

C. Whole genome screens

Another approach is to screen the whole human genome for linkage with a disease without any assumptions on disease pathogenesis (whole genome screening) (59). Whole genome screening is performed by testing a panel of markers that span the entire human genome for linkage with a disease in a given dataset. If one or more of the markers shows evidence for linkage with the disease according to the guidelines of Lander and Kruglyak (66), these regions may harbor susceptibility genes for the disease studied. These linked regions can then be fine-mapped, and the genes can be identified (see Section III.E). The two requirements for performing parametric linkage from a whole genome screen in a complex disease are: 1) the availability of a sizeable and well-validated dataset of multiplex families (large families with more than one individual affected); and 2) the availability of a map of highly polymorphic, closely-spaced markers covering the whole genome, which can be tested for linkage with the disease.

D. Markers used in linkage and association studies

1. Microsatellites. The most widely used polymorphic markers for whole genome screening are microsatellite markers. Mi-

microsatellites are regions in the genome that are composed of short sequence repeats, most commonly two-base CA-repeats (81) and usually have no known physiological function. Microsatellite loci are highly polymorphic (*i.e.*, have many alleles) because the number of repeats in each individual is variable. Moreover, they are extremely abundant and uniformly distributed throughout the genome at distances of less than 1 million bp (81). Therefore, microsatellites serve as excellent markers in whole genome linkage studies. Two independent whole genome screens have now been completed for the AITDs (see *Section VII*). Microsatellites can also be useful in candidate gene studies if a microsatellite is identified within the candidate gene or very close to it.

2. Single nucleotide polymorphisms (SNPs). SNPs are single base pair positions in genomic DNA at which different sequence alternatives (alleles) exist in normal individuals. Although four alleles are theoretically possible (A, C, T, G), in humans, most SNPs are diallelic (~70% are A/G; reviewed in Ref. 82). SNPs are very abundant, and current data suggest that their frequency is about one SNP per 1000 bp (83). Because SNPs are less informative than microsatellites (SNPs have only two alleles, and microsatellites usually have more than five alleles), they are not useful in linkage studies. However, because SNPs are much more abundant and closely spaced than microsatellites, they are ideal for fine-mapping genes in linked regions using association studies. The importance of SNPs stems from the fact that many have the potential to change the amino acid sequence of a gene product and be directly involved in the susceptibility to complex diseases because they cause changes in gene function (82). Thus, if a SNP allele inside a gene is found to be significantly associated with a disease, it may be the actual causative allele, increasing susceptibility to the disease (84).

E. A suggested algorithm for searching for complex disease genes

Unlike the search for genes causing simple Mendelian disorders, it is still not known what is the best approach to identify susceptibility genes for complex diseases. However, several groups have successfully identified putative susceptibility genes in complex disorders, notably the identification of NOD2 as a major gene for Crohn's disease (60, 84), and the approach used in these studies is gaining popularity (85, 105). It is based on five steps that combine genetic linkage and association analyses with functional-biological studies:

- 1. Identifying linked loci.** This is achieved by whole genome screening using microsatellite markers at an average distance of less than 10 cM.
- 2. Confirming linked loci.** A linked locus should be confirmed by finding evidence for linkage in two independent datasets (66). Confirmed loci most likely harbor susceptibility genes, *e.g.*, HLA in type 1 diabetes mellitus (DM) (59).
- 3. Fine-mapping confirmed loci.** Several methods are available to fine-map a linked region. The most popular approach for fine-mapping loci linked with complex disease is LD mapping. LD mapping is based on association studies with markers that saturate the region of interest. The marker that shows

the strongest association with the disease (*i.e.*, the lowest *P* value and highest RR) is probably closest to the disease gene. This method can narrow down the region of interest to a few hundred kilobases. LD mapping may be limited in populations without abundant founder haplotypes. However, in carefully selected populations with homogenous disease etiology, it may be very effective (64, 83). In addition, this method is effective in regions showing strong evidence for linkage with the disease (83).

4. Testing genes in the linked region. After the linked region has been narrowed to less than 1 cM, the known and unknown genes in this region can be analyzed. The Human Genome Project identified most of the genes in a linked region, although it is still possible that an unidentified gene may not be annotated in the Human Genome Project maps. Known genes can be analyzed by identifying SNPs in them and testing these SNPs for association with the disease. If a certain SNP shows a consistently significant association with the disease, it may be the susceptibility allele in the region, although LD with another disease-causing polymorphism cannot be ruled out.

5. Functional studies. To demonstrate that an associated allele is a true susceptibility allele, it is necessary to show that it affects the function of the gene in a way that increases the risk of developing disease. This provides indirect evidence that it may be the actual susceptibility allele for the disease. For example, the SNP in the NOD2 gene that is associated with Crohn's disease was shown to influence the activation of nuclear factor- κ B, thereby influencing the activation of intestinal T cells (84). Although there is no absolute way to prove that a certain SNP is the true susceptibility allele for a disease, as the function of the SNP and its influence on disease development is clarified the role of the identified SNP can be substantiated.

IV. Immune Regulatory Genes Studied in AITD

A. The role of HLA in the genetic susceptibility to AITD

1. The MHC. The MHC region, encoding the HLA glycoproteins, consists of a complex of genes located on chromosome 6p21. The MHC region also encodes various additional proteins, most of which are associated with immune responsiveness (86). The MHC locus itself encodes genes that are grouped into three classes: 1) class I genes, including the HLA antigens A, B, and C; 2) class II genes, including the HLA-DR, DP, and DQ genes; and 3) class 3 genes, including several complement components (*e.g.*, C4), TNF α , heat shock protein 70, and several other genes (for a review, see Ref. 87). Because the HLA region is highly polymorphic and contains many immune response genes, it was the first candidate genetic region to be studied for association and linkage with AITD.

2. Association of HLA with GD. GD was initially found to be associated with HLA-B8 in Caucasians (88) (Table 3). In these early studies, HLA-B8 was associated with RRs for GD ranging from 1.5–3.5 (89). Subsequently, it was found that GD was more strongly associated with HLA-DR3, which is now

known to be in LD with HLA-B8 (Table 3; reviewed in Ref. 90). The frequency of DR3 was generally 40–55% in GD patients and approximately 15–30% in the general population, giving a RR for people with HLA-DR3 of up to 4.0 (Table 3) (89, 91, 92; reviewed in Ref. 93). A recent family-based study from the United Kingdom using TDT analysis con-

firmed the results of the case control studies (94). The HLA genes were shown to be associated with GD in non-Caucasians, as well, although the associated alleles were different (Table 4). Studies in the Japanese population have shown associations of GD with HLA-B35 (95, 96). However, other HLA alleles have also been reported to be increased in Japanese GD patients (97–99). In the Chinese population, an increased frequency of HLA-Bw46 has been reported (100–104), and in African-Americans an increased frequency of HLA DRB3*0202 was reported (Table 4) (302). Interestingly, one study of a mixed population in Brazil showed association with HLA-DR3, implying that this allele may confer susceptibility in other ethnic groups and not just Caucasians (106). Alternatively, this Brazilian population was mostly of European ancestry. Among Caucasians, HLA-DQA1*0501 was also shown to be associated with GD (RR, 3.8; Table 3) (Refs. 107–109), but recent studies have suggested that the primary susceptibility allele in GD is indeed HLA-DR3 (HLA-DRB1*03) (110). HLA-DR3 has over 20 subtypes (111). We recently subtyped, by direct sequencing, HLA-DR3 in a population of DR3-positive GD patients and controls (112). The allelic frequency of DRB1*0311 was significantly lower in patients than in controls (112). These results suggested that GD is associated with specific sequences of the DR3 allele. The pattern of transmission of HLA alleles from parents to offspring was also studied. A recent study suggested a preferential transmission of HLA susceptibility alleles from fathers to affected offspring, whereas maternal susceptibility alleles were not transmitted more frequently than expected (113). This may suggest parental imprinting in the transmission of HLA susceptibility alleles to affected offspring.

The role of HLA polymorphisms in the clinical expression of GD has also been explored. Some groups reported an association between the likelihood of relapse of GD and

TABLE 3. Some of the important HLA association studies in GD performed in Caucasians

Country	No. of patients	HLA allele	RR	Ref.
Belgium	194	DRB1*0301	2.53	110
Canada	175	B8	3.1	89
		DR3	5.7	
Denmark	86	B8	2.80	88
		Dw3	3.94	
France	94	B8	3.4	117
		DR3	4.2	
Germany	253	DR3	2.52	114
Hungary	256	B8	3.48	290
		DR3	4.8	
Ireland	86	B8	2.5	116
		DR3	2.6	
Canada	133	DR3	2.63	291
Sweden	78	B8	4.4	115
		DR3	3.9	
United Kingdom	127	B8	2.77	118
		DR3	2.13	
United Kingdom	65	DR3	3.20	292
United Kingdom	101	DR3	1.10	293
United States	65	DR3	3.38	92
United States	92	DRB1*03	2.6	294
		DRB1*08	3.2	
United States	60	DR3	3.4	124
United Kingdom	120	DQA1*0501	3.8	107
United Kingdom	228	DRB1*0304	2.7	94
		DQB1*0301	1.9	
		DQA1*0501	3.2	
United States	94	DQA1*0501	3.71	108

TABLE 4. Major HLA association studies in GD performed in non-Caucasian populations

Country	Ethnic group	No. of patients	HLA allele(s)	RR/P value	Ref.
Hong Kong	Chinese	132	Bw46	4.8	295
Hong Kong	Chinese	67 (children)	DQB1*0303	4.2	296
Hong Kong	Chinese	97	B46	2.3	101
			DR9	2.2	
			DQB1*0303	3.2	
Singapore	Chinese	35	B46	8.2	102
Singapore	Chinese	159	Bw46	4.2	104
Japan	Japanese	33	Bw35	$P < 0.02$	95
Japan	Japanese	106	B46	$P < 0.0004$	97
Japan	Japanese	30	DR5	8.14	297
			DRw8	3.07	
Japan	Japanese	62	DRB1*0803	$P < 0.02$	98
Japan	Japanese	76	A2	2.86	298
			DPB1*0501	5.32	
Korea	Korean	128	B13	3.8	299
			DR5	4.4	
			DRw8	2.3	
India	Asian Indian	57	B8	4.1	300
			DQw2	5.4	
United States	African-American	73	No association		301
United States	African-American	49	DRB3*0202	3.6	302
South Africa	African	103	DR1	3.5	303
			DR3	2.4	
Brazil	Mixed	75	DRB1*0301	2.8	106
			DQA1*0501	3.74	

HLA-DR3, but most other investigators were unable to confirm this observation (114–116). Studies of HLA associations in Graves' ophthalmopathy (GO) have produced conflicting results, with some workers reporting increased frequency of HLA-DR3 in patients with GO, and others reporting no difference in the distribution of HLA-DR alleles between GD patients with and without ophthalmopathy (88, 89, 117, 118). These results were not surprising in view of our recent segregation analysis that showed no genetic influences on the development of GO (41). Likewise, no difference in the DR3 frequency was found in GD patients with and without pretibial myxedema (89). Some workers have suggested that local factors such as orbital pressure play an important role in the development of GO and pretibial myxedema (119).

3. *Association of HLA with HT.* Data on HLA haplotypes in HT have been less definitive than in GD. A general methodological problem has been disease definition. Although the diagnosis of GD may be relatively straightforward, the definition of HT has been more controversial. HT encompasses a spectrum of manifestations, ranging from the simple presence of thyroid autoantibodies with focal lymphocytic infiltration, which may be of no functional consequence (asymptomatic autoimmune thyroiditis), to the presence of goitrous or atrophic thyroiditis, characterized by gross thyroid failure (31). Initial studies failed to demonstrate an association between goitrous HT and HLA A-, B-, or C-antigens (120). Later studies showed an association of goitrous HT with HLA DR5 (RR, 3.1) (121) and of atrophic HT with DR3 (RR, 5.1) (122) (Table 5). Association of HT with HLA DR3 in Caucasians has been confirmed in subsequent studies (123, 124) and further supported by studies of transgenic mice (125). An association between HT and HLA-DQw7 (DQB1*0301) has also been reported in Caucasians (126, 127). In non-Caucasian ethnic groups, different HLA haplotypes were reported to be associated with HT, e.g., HLA-DRw53 in Japanese (128) and HLA-DR9 in Chinese (129) (Table 5).

4. *HLA linkage studies in AITD.* Linkage studies of HLA in AITD have been largely negative (130–134). Only one recent study from the United Kingdom showed weak evidence for linkage between GD and the HLA region (135), and an additional study reported linkage only when conditioning on

DR3 (136). It is difficult to explain why the HLA genes show consistent association with GD but no evidence for linkage. The lack of linkage means that HLA-DR3, as measured, did not cause the familial segregation of GD, whereas the relatively strong association showed that HLA-DR3 conferred a generalized increase in risk for GD in the general population. Indeed, we were able to show that HLA was associated with GD in both sporadic GD patients and probands from GD families, giving similar RRs (our unpublished data).

B. The role of CTLA-4 in the genetic susceptibility to AITD

1. *The cytotoxic T lymphocyte antigen-4 (CTLA-4) costimulatory molecule.* CTLA-4 is an important costimulatory molecule that participates in the interaction between T cells and antigen-presenting cells (APC). APC activate T cells by presenting to the T cell receptor an antigenic peptide bound to an HLA class II protein on the cell surface. However, a second signal is also required for T cell activation, and these costimulatory signals may be provided by the APC themselves or other local cells (137). The costimulatory signals are provided by a variety of proteins that are expressed on APC (e.g., B7-1, B7-2, B7h, CD40) and interact with receptors (CD28, CTLA-4, and CD40L) on the surface of CD4+ T lymphocytes during antigen presentation (137) (Fig. 3). Whereas the binding of B7 to CD28 on T cells costimulates T cell activation, the presence of CTLA-4, which has a higher affinity for B7, down-regulates T cell activation by competing for the binding of B7 to CD28. The suppressive effects of CTLA-4 on T cell activation have raised the possibility that mutations altering CTLA-4 expression and/or function could result in an exaggerated T cell activation and lead to the development of autoimmunity.

2. *CTLA-4 association studies in GD.* Recently, there have been several reports demonstrating an association between the CTLA-4 gene and the AITDs (Table 6) (138–142). The initial studies found an association between a microsatellite marker located at the 3' untranslated region (UTR) of the CTLA-4 gene and GD, giving a RR of 2.1–2.8 (138, 141). Later, two SNPs were also identified in the CTLA-4 gene: 1) at position 49 in the CTLA-4 leader peptide (A/G₄₉), resulting in an alanine/threonine polymorphism; and 2) in the promoter of

TABLE 5. Selected HLA association studies in HT

Country	Ethnic group	No. of patients	HLA allele	RR/P value	Ref.
Canada and United Kingdom	Caucasians	64	DR4 DR5 DQw7	2.9 3.8 4.7	127
Italy	Caucasians	126	DRB1*04- DQB1*0301	$P < 0.05$	156
Canada	Caucasians	40	DRw3	3.5	122
Canada	Caucasians	40	DR5	3.1	121
United Kingdom	Caucasians	49	DQB1*0301 DQA1*0301/2	Not reported	126
United Kingdom	Caucasians	36	DR5 DQ7	3.5	305
United Kingdom	Caucasians	86	DR3	2.23	123
China	Chinese	53	DRw9	$P < 0.05$	129
Japan	Japanese	99	DRw53	3.33	128
Japan	Japanese	50	Bw46 B51	3.66 3.42	306

CTLA-4 at position -318 (C/T₋₃₁₈). Case-control studies from several groups, including our own group, have shown an association between the alanine (G) polymorphism and GD with a RR of approximately 2.0 (Table 6) (41, 140, 143–145). The association between GD and the CTLA-4 3' UTR microsatellite and A/G₄₉ SNP has been consistent across populations of different ethnic backgrounds such as Caucasians (138), Japanese (145, 146), and Koreans (147). The association of CTLA-4 and GD has also been confirmed in a family-based study using TDT analysis (148). In contrast, association studies using the C/T₋₃₁₈ SNP of CTLA-4 have been less consistent, with some showing association (144) and others not (149).

3. *CTLA-4 association studies in HT.* Because CTLA-4 is a nonspecific costimulatory molecule, it is expected to confer

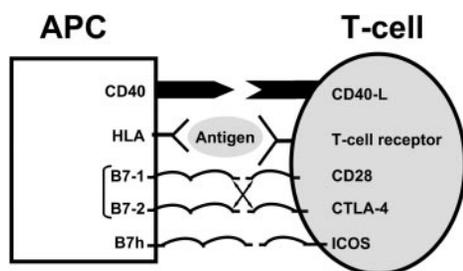


FIG. 3. T cell activation by APC. The APC presents a peptide antigen bound to HLA class II molecules, and the peptide is recognized by the T cell receptor. This interaction will not lead to T cell activation without additional costimulation provided by costimulatory molecules. Engagement of B7 molecules with CD28 provides costimulation, whereas CTLA-4 binding to the B7 molecules blocks CD28 activation by B7. In addition, CTLA-4 engagement with B7 directly suppresses T cell activation. Additional costimulation is provided by activation of inducible costimulator by its ligand (B7h), and by CD40-CD40L (CD40L or CD154) interaction. CD40 is also expressed on B cells, and its activation results in B cell differentiation, Ig production, and isotype switching.

TABLE 6. Major CTLA-4 association studies in AITD

CTLA-4 polymorphism	Country	Ethnic group	Disease	No. of patients	RR/P value	Ref.
CTLA-4(AT)	United States	Caucasians	GD	133	2.82	138
CTLA-4(AT)	United Kingdom	Caucasians	GD	112	2.1	141
			HT	44	2.2	
CTLA-4(AT)	Hong Kong	Chinese	GD	94	$P = 0.037$	139
CTLA-4(AT)	Japan	Japanese	GD	62	NS	155
			HT	30	NS	
			GD+HT	349	1.8	146
Thr/Ala (A/G) ₄₉	Germany	Caucasians	GD	305	2.0	140
Thr/Ala (A/G) ₄₉	United Kingdom	Caucasians	GD	94	$P = 0.003$	163
Thr/Ala (A/G) ₄₉	United Kingdom	Caucasians	GD	379	1.6	148
Thr/Ala (A/G) ₄₉	United Kingdom	Caucasians	GD	484	$P < 0.0001$	162
Thr/Ala (A/G) ₄₉	United States	Caucasians	GD	85	1.6	41
Thr/Ala (A/G) ₄₉	Germany	Caucasians	HT	73	$P < 0.04$	154
Thr/Ala (A/G) ₄₉	Italy	Caucasians	HT	126	NS	156
Thr/Ala (A/G) ₄₉	United Kingdom	Caucasians	HT	158	1.57	143
Thr/Ala (A/G) ₄₉	Slovenia	Caucasians	Tab's	67	$P < 0.005$	158
Thr/Ala (A/G) ₄₉	Japan	Japanese	GD	153	2.64	145
Thr/Ala (A/G) ₄₉	Korea	Korean	GD	97	1.6	147
			HT	110	NS	
C/T (-318)	United Kingdom	Caucasians	GD	188	NS	149
	United Kingdom	Caucasians	HT	90	NS	
	Hong Kong	Chinese	GD	98	NS	
C/T (-318)	Germany	Caucasians	GD	125	$P = 0.006$	144

NS, Not significant.

susceptibility to AITD and autoimmunity in general and not specifically to GD (150). Indeed, CTLA-4 was reported to be associated and linked with all forms of AITD (GD, HT, and Tab; see Section IV.B.4), and with many autoimmune diseases such as type 1 diabetes (139, 140, 151), Addison's disease (152), and myasthenia gravis (153). CTLA-4 has been reported to be associated with HT in various populations including Caucasians (141, 143, 154) and Japanese (146, 155). There have also been two reports of no association of HT with CTLA-4, most likely due to lack of power (144, 156).

4. *CTLA-4 and Tab.* Two studies have now shown that CTLA-4 confers susceptibility to the production of thyroid antibodies. Our group has shown strong evidence for linkage between the CTLA-4 gene region and the production of thyroid antibodies with a MLS of 4.2 (157). Recently, another report has described an association between the G allele of the CTLA-4 A/G₄₉ SNP and the thyroid autoantibody diathesis (158). Because the development of thyroid antibodies often represents the preclinical stage of AITD (159), it is possible that CTLA-4 predisposes, nonspecifically, to the development of thyroid autoimmunity. Additional genetic and/or environmental factors must be necessary for the development of the specific GD/HT phenotypes (150).

5. *CTLA-4 and disease severity.* Several studies have examined whether CTLA-4 polymorphisms influence disease severity. Heward *et al.* (148) reported that the CTLA-4 A/G₄₉ SNP G allele was associated with more severe thyrotoxicosis at diagnosis (as reflected by higher free T₄ levels). Similar findings were reported by Park *et al.* (147) but not by Zaletel *et al.* (158). It has also been reported that the frequency of the G allele and the GG genotype of the CTLA-4 A/G₄₉ SNP was significantly higher in GD patients who did not go into remission after 5 yr on antithyroid medications (160). In addition, CTLA-4 has been shown to be associated with GD in children (161).

Together, these studies may suggest that CTLA-4 may influence both the initiation of AITD and the severity of the phenotype.

CTLA-4 polymorphisms have also been tested for association with GO. Most studies have been negative and did not show that CTLA-4 conferred a specific risk for GO beyond that conferred for GD (41, 158, 162). However, two groups have reported an association between GO and CTLA4 (147, 163). It is most likely that the reported CTLA-4 associations with GO reflected an association between CTLA-4 and more severe GD and not a specific association with the GO phenotype. Indeed, our recent segregation analysis showed no evidence for any genetic susceptibility to GO (41).

6. Family linkage studies. Vaidya *et al.* (135) reported linkage to the CTLA-4 gene region on chromosome 2q33 in families with GD using nonparametric linkage analysis. The linkage became stronger when families with AITD, rather than just GD, were included in the study, again demonstrating that CTLA-4 most likely confers general susceptibility to thyroid autoimmunity and not to a specific AITD phenotype. As discussed earlier, and in keeping with the view that the CTLA-4 gene predisposes to thyroid autoimmunity rather than to one specific disease, we found strong linkage between the CTLA-4 gene region and TAb (157). However, the region on chromosome 2q33 containing the CTLA-4 gene is replete with candidate immune regulatory genes for thyroid autoimmunity (*e.g.*, CD28 and inducible costimulator), and it was unclear whether the CTLA-4 gene itself or another immune regulatory gene in the region was involved in the genetic susceptibility to AITD. Recently, we and others tested additional genes and markers in the 2q33 region, and the strongest association was with the CTLA-4 markers (151, 157). These results were in keeping with results obtained in type 1 DM (164, 165).

C. Other immune-related genes

1. T cell receptor genes. The initiation of GD involves infiltration of the thyroid by T cells reactive with thyroid antigens. These T cells are oligoclonal and are restricted by their T cell receptor V gene use (166). Thus, the T cell receptor seemed a likely candidate gene for thyroid autoimmunity and has, therefore, been studied extensively. Earlier studies reported an association between a restriction fragment length polymorphism of the T cell receptor β -chain and GD (167). However, these results were not confirmed by other groups (92, 168). We have also studied the T cell receptor α -genes (on chromosome 14) and β -genes (on chromosome 7) by linkage studies using microsatellite markers located at or close to these genes (133). These markers gave highly negative LOD scores with GD, HT, and AITD, thus ruling out linkage of the T cell receptor α - and β -genes to AITD (133). Therefore, it can be concluded that the T cell receptor genes are not major susceptibility genes for AITD.

2. The IgG heavy chain (IgH) gene. Another good candidate immune regulatory gene for GD was the IgH gene, because the hallmark of GD is the production of stimulating TSHR IgG autoantibodies. Indeed, the IgH gene was one of the first genes studied in AITD. Early studies reported an association

between IgH Gm allotypes and AITD in the Japanese population (169–171). However, these results have not been reproduced in other populations (172, 173). Similarly, we were unable to confirm linkage between the IgH gene and GD in a Caucasian dataset of families (174).

3. Immune regulatory genes. Other genes tested for association with GD include the IL-1 receptor antagonist gene (175–178), TNF α gene (133, 179), interferon- γ gene (180), and the transporters associated with antigen presentation genes (181). However, none of these produced replicable associations with GD. Recently, an association between a promoter polymorphism of the IL-4 gene and GD has been reported, but a subsequent study could not replicate these results (182). The vitamin D receptor, which may have some immune modulatory functions, has also been reported to be associated with GD (183), and another study reported an association of GD with a vitamin D binding protein (184). These results need to be confirmed, and it cannot be excluded that other genes in LD with these genes are the susceptibility genes at these loci.

V. Thyroid-Specific Genes Studied in AITD

A. TSH receptor

The hallmark of GD is the production of the TSHR antibodies. Therefore, the TSHR gene was long thought to be a likely candidate gene for GD. Three common germline SNPs of the TSHR have been described (185). Two of them are located in the extracellular domain of the TSHR: an aspartic acid to histidine substitution at position 36 (D36H), and a proline to threonine substitution at position 52 (P52T). The third SNP is a substitution of glutamic acid for aspartic acid (D727E) within the intracellular domain of the receptor. Most studies on the contribution of the TSHR gene to the genetic susceptibility to GD have focused on the SNPs in the extracellular domain of the TSHR (186–191) because this domain is responsible for TSH and TSHR antibody binding. Amino acid changes in the extracellular domain of the TSHR could theoretically change the amino acid sequence of TSHR T cell epitopes (192). Initial studies suggested that the P52T SNP was associated with GD in females (186). However, other authors were unable to confirm the association between the P52T SNP and GD in Caucasians (187–191). The D36H SNP has also been reported not to be associated with GD (189). Linkage studies in GD families using three microsatellite markers within introns 2 and 7 of the TSHR gene have also been negative in Caucasians (193, 194). Association studies using these markers also have been negative in Caucasians (41). However, an association between AITD and TSHR microsatellite markers has been reported in the Japanese (146, 155), and recently, the D727E SNP was reported to be associated with GD in a Caucasian Russian population (195), but these results were not replicated in a subsequent study by another group (196). We recently tested whether the D727E SNP was associated with GD in general and with disease severity. The results of our study did not show an association between the D727E SNP and GD and did not show an effect of the D727E SNP on the GD phenotype or disease severity (197). The frequency of the G allele was not increased in

patients with more severe forms of GD (*i.e.*, ophthalmopathy and goiter) and in patients with early disease onset. However, our study and other negative TSHR studies could not exclude a weak association between GD and the TSHR gene because very large datasets are needed to detect associations with low RRs. We, therefore, performed a meta-analysis combining our data with the data reported in the previous two negative TSHR studies. The results of the meta-analysis showed a weak association between the D727E SNP E allele and GD ($P = 0.03$; RR = 1.6) (197). Therefore, it remains possible that the TSHR is a minor susceptibility gene for GD.

B. Thyroid peroxidase

Antibodies to TPO are the most specific marker for HT, and therefore, TPO is another possible candidate gene for AITD. It was recently found that autoantibodies recognizing immunodominant epitopes of TPO are genetically transmitted within families (50). One possible explanation of the genetic transmission of TPO-Ab epitopes is that this is caused by changes in TPO gene sequence and/or expression in the thyroid. The TPO gene was tested for linkage and association with AITD in two studies using a microsatellite inside the TPO gene. However, these studies showed no evidence of linkage and/or association of the TPO gene with AITD (174, 198). Therefore, the TPO gene does not seem to be a major susceptibility gene for AITD, although a minor role cannot be excluded.

C. Thyroglobulin

1. *Tg plays a central role in the development of thyroid autoimmunity.* Tg is the major protein product synthesized in the thyroid gland, and Tg autoantibodies are common in AITD (reviewed in Ref. 199). There is abundant, indirect, evidence that Tg plays an important role in the etiology of AITD including: 1) anti-Tg-Ab are detected in almost all patients with AITD, both GD and HT (200, 201), and there is evidence that Tg-Ab of AITD patients are restricted in their epitope specificity in contrast to the polyclonal nature of Tg-Ab found in healthy individuals (202); 2) immunization with Tg induces autoimmune thyroiditis in experimental animals (203), and the induction of experimental autoimmune thyroiditis (EAT) in mice by Tg is well known to be MHC-dependent (204, 205); this last observation implies an interaction between the Tg molecule and the MHC glycoproteins in the induction of thyroiditis; 3) Tg peptides containing the hormonogenic sites at positions 5, 2553, and 2567 (206–208), and other specific Tg peptides (209, 210) can, on their own, induce EAT in susceptible mice, implying that specific epitopes on Tg can stimulate the development of AITD; 4) spontaneous models of autoimmune thyroiditis in the NOD mouse and BB/W rat are also characterized by the development of anti-Tg-Ab (211, 212); and 5) HLA-DR3 transgenic mice that were murine class II-negative developed EAT after immunization with human Tg (125), showing that human HLA-DR3 could present conserved Tg epitopes in originally thyroiditis-resistant mice. Moreover, it has been shown that the serum levels of Tg are genetically determined and may influence the susceptibility to AITD (213).

2. *The Tg gene region is linked with AITD.* Two whole genome screens have shown strong evidence for linkage of AITD with the chromosome 8q24 region, which contains the Tg gene (134, 214) (see Section VII). Because the Tg gene was located within a linked region, we proceeded to analyze the Tg gene directly. We identified two new Tg microsatellites in intron 10 (designated Tgms1) and intron 27 (designated Tgms2). Linkage analysis using Tgms2 gave a two-point LOD score of 2.1 and multipoint LOD score of 2.9, confirming that the Tg gene was linked with AITD (214).

3. *The Tg gene is also associated with AITD.* We then used the same two Tg microsatellites to test whether the Tg gene was associated as well as linked with AITD. Using an unselected group of 190 Caucasian GD patients and 134 age- and sex-matched Caucasian controls, we found a weak association between Tgms2 and AITD ($P = 0.05$; RR = 1.4) (214). However, the association was stronger when just the probands from the linked families ($n = 32$) were used ($P = 0.004$; RR = 2.3). TDT analysis also showed an association of Tgms2 with AITD ($P = 0.02$), although with a different allele, suggesting that Tgms2 is in LD with another polymorphism of the Tg gene. Thus, the Tg gene was both linked and associated with AITD and, therefore, could be an important AITD susceptibility gene. However, it remains possible that another gene on 8q24 in LD with Tg was the AITD susceptibility gene responsible for the observed linkage and association at this locus.

VI. Linkage Studies of Candidate Chromosomal Regions (Table 7)

A. Chromosome X

The AITDs are 5–10 times more common in women (159). The increased female preponderance of AITD may potentially be explained by the effects of estrogenic sex steroids in promoting autoimmunity (215), by genetic factors, or as a consequence of pregnancy and the resulting maternal microchimerism (216). Although there are studies supporting a role for estrogen in the induction of autoimmunity (215, 217–220) these results have not been consistent, and some studies have even shown suppression of thyroiditis (221) and other autoimmune diseases (222) by estrogens. However, there are also indirect data suggesting that chromosome X abnormalities might be responsible for the increased incidence of AITD in women. These data come mostly from studies of patients with Turner's syndrome. There is a strong association of Turner's syndrome with the production of thyroid autoantibodies and autoimmune thyroiditis (223–225). Up to 50% of patients with Turner's syndrome develop thyroid antibodies in early childhood (224, 225), and up to 20% develop clinical disease (223, 225). Studies on the correlation between the karyotype and AITD have shown that 83% of patients with Turner's syndrome who had X-isochromosomes developed TAb, and 57% developed clinical AITD (223, 226). These results suggested that a gene on chromosome Xq may play a role in the development of AITD (226). Two studies have subsequently examined the X chromosome for linkage with AITD. We showed evidence for

TABLE 7. Ten loci reported to show evidence for linkage with AITD

Chromosome	Locus	Marker	Phenotype	Type of study	Type of analysis	Results	Ref.
2	2q33 (CTLA-4)	D2S155	TAbs	WGS	Ped-LOD	HLOD = 4.2	157
2	2q33 (CTLA-4)	D2S117	GD	CLA	ASP-NPL	NPL = 3.4	135
5	5q31	D5S356	AITD	WGS	ASP-LOD	LOD = 3.1	134
6	6p21 (HLA)	D6S273	GD	CLA	ASP-NPL	NPL = 1.95	135
6	6p11 (AITD-1)	D6S257	AITD	WGS	Ped-LOD	LOD = 2.9	193
6	6p11	D6S257	HT	WGS	One large pedigree	LOD = 1.52 NPL = 7.53	307
8	8q24 (Tg)	D8S272	AITD	WGS	ASP-LOD	LOD = 2.3	134
8	8q24 (Tg)	D8S284	AITD	WGS	Ped-LOD	HLOD = 3.5	228
12	12q22 (HT-2)	D12S351	HT	WGS	Ped-LOD	HLOD = 2.3	193
14	14q31	D14S81	GD	WGS	Ped-LOD	LOD = 2.1	193
18	18q21 (IDDM-6)	D18S487	GD	CLA	ASP-NPL	NPL = 3.1	247
20	20q11 (GD-2)	D20S195	GD	WGS	Ped-LOD	LOD = 3.5	193
20	20q11 (GD-2)	D20S106	GD	CLA	ASP-NPL	NPL = 2.01	251
X	Xp11 (IDDMX)	DXS8083	GD	CLA	ASP-NPL	NPL = 2.01	229

ASP, Affected sib-pairs; CLA, candidate locus analysis; LOD, LOD score analysis; NPL, nonparametric linkage analysis; Ped, multiplex multigenerational pedigrees; WGS, whole genome screening.

linkage at chromosome Xq21 (227), but this locus was not confirmed in our second dataset (228). Another study showed evidence for linkage at Xp11 with a maximum non-parametric LOD score of 2.2 (229) (Table 7). More studies of the X chromosome are needed to confirm whether a gene on the X chromosome confers susceptibility to AITD.

There are a number of possible mechanisms whereby the X chromosome could influence the development of AITD. Females have two X chromosomes (one paternal and one maternal), whereas males have only one X chromosome (maternal). Therefore, females are twice as likely to inherit an X chromosome AITD susceptibility gene as males. Several immune regulatory genes are located on the X chromosome (e.g., the CD40 ligand gene), but their involvement in autoimmunity has not been studied. Another possible mechanism is through X-inactivation. X-inactivation in females results in the production of two classes of cells that differ in the transcription of X chromosome-encoded genes, including genes coding for self-antigens. If these two cell classes extend to the thymic cells responsible for tolerizing T cells in embryonic life, the immune repertoire will not be entirely tolerized to one version of the two self-antigens encoded by the X chromosome. Such lymphocytes would be autoreactive to that antigen and could induce an autoimmune response (230). Indeed, some workers have suggested that skewed X-chromosome inactivation in the thymus may lead to inadequate thymic deletion and autoimmunity (231). Although this is an attractive theory that could help explain the female preponderance of autoimmune conditions (because this escape mechanism from tolerance can occur only in females), there are no data to support it.

B. The IDDM loci

Type 1 diabetes [insulin-dependent DM (IDDM)] is known to be associated with AITD (232–235; reviewed in Ref. 236). Up to 20% of patients with IDDM have thyroid antibodies (237), and 50% of IDDM patients with TAb will develop clinical AITD (238). In addition, postpartum thyroiditis is twice as common in women with IDDM (239–241). Therefore, it is possible that AITD and IDDM share genetic sus-

ceptibility. Moreover, a recent study has shown that 35.9% of IDDM probands from the Familial Autoimmune Diabetes Study had AITD (26.6% had HT and 9.3% had GD) (242). These results implied that the association between IDDM and AITD is even stronger in cases of familial IDDM and may point to common susceptibility genes. To date, more than 15 type 1 diabetes loci have been suggested (59, 85, 139, 243–245), and some of these sites have also been evaluated in patients with AITD. One locus that has been found to be shared by type 1 diabetes and AITD is the CTLA-4 gene region discussed earlier. This genetic region is linked with both type 1 diabetes (139) and AITD (135). Moreover, in both type 1 diabetes and AITD it was shown that the CTLA-4 gene is most likely the susceptibility gene at this locus (157, 164, 165), although the specific polymorphism of CTLA-4 responsible for this association is not known. Other IDDM loci have also been tested in AITD. IDDM-2 on chromosome 11p, IDDM-4 on chromosome 11q, IDDM-5 on chromosome 6q25, and IDDM-8 on chromosome 6q27 did not show evidence for linkage and/or association with AITD (174, 246, 247). On the other hand, a study from the United Kingdom showed evidence for linkage of IDDM-6 on chromosome 18q with AITD (247) (Table 7), although we did not find this site linked to AITD in our own whole genome screening.

C. The 14q region

The chromosome 14q region caught the attention of several investigators because it harbors the TSHR gene, and therefore, it has been tested for linkage with AITD. In our preliminary screen of the 14q region using 56 multiplex families, we obtained evidence for linkage with GD at this locus (designated GD-1) with an MLS of 2.5 (174, 248). This locus was distinct from the TSHR gene, which did not show strong evidence for linkage with GD (248). We have recently expanded our dataset to include 102 families (540 individuals) and were able to replicate the evidence for linkage at the 14q region (228). Imrie *et al.* (229) also tested the 14q region and reported no evidence for linkage at GD-1. However, they tested only a few markers in a very narrow region that did not include the area of MLS obtained in our expanded data-

set. Thus, additional linkage studies are needed to confirm the evidence for linkage at this locus. The GD-1 locus contains several potential positional candidate genes. One of them is a newly isolated growth factor, suppressor of lin-12-like protein (249), which was tested, but no association was found with GD (250). Another interesting positional candidate gene in this locus is the TGF β 3 gene.

D. The 20q region

Chromosome 20 was one of the first chromosomes tested in our preliminary genome scan. Because it showed strong evidence for linkage at 20q, this region was studied in more detail. Using a dataset of 56 multiplex families, we identified a locus on chromosome 20q11 showing strong evidence for linkage with GD with a MLS of 3.5 (79, 193). This GD locus was not linked to HT, because analysis of the data for the HT families gave strongly negative LOD scores. Moreover, in families with GD- and HT-affected individuals, the locus was linked only with GD, demonstrating its high specificity for GD. This region was recently tested by another group from the United Kingdom. Although this locus did not show evidence for linkage in the whole United Kingdom dataset, subsetting the families by transmission showed evidence for linkage with GD in the families with vertical transmission (consistent with dominant complex inheritance) of disease (251). This further supported the 20q11 locus as an important susceptibility locus for GD.

Recently, we have expanded our dataset to 102 multiplex families and used this expanded dataset to fine-map the 20q11 locus using 10 densely spaced microsatellite markers. Linkage analysis in our expanded dataset gave a MLS of 1.2 with heterogeneity. However, a subset of the families (Caucasian families of non-Italian origin) gave a MLS (without heterogeneity) of 3.3 (252). Thus, the 20q region was linked with a subset of our dataset (252) and a subset of the United Kingdom dataset (251). It is still unclear what is unique to these subsets, but they may represent a different phenotype of GD. The CD40 gene, an important immune modulator, is located within the linked region on chromosome 20q11, and therefore, it was a likely positional candidate gene for GD. CD40 is a transmembrane glycoprotein that is expressed predominantly on B cells, but also on monocytes, dendritic cells, epithelial cells, and other cells (reviewed in Ref. 253). It is a member of the TNF receptor superfamily, and it binds to a ligand (CD40L or CD 154) that is expressed mainly on activated T cells. Binding of CD40L to CD40 induces B cells to proliferate and undergo Ig isotype switching (254). CD40 has been shown to play an essential role in the regulation of humoral immunity, central and peripheral T cell tolerance, and APC function (reviewed in Ref. 255). Moreover, *in vivo* blockade of CD40 has been shown to suppress the induction of EAT (256). Therefore, we tested whether CD40 was the GD susceptibility gene on chromosome 20q11. Sequencing of the CD40 gene revealed a C/T SNP in the Kozak sequence of the gene. Analysis of the CD40 Kozak SNP in 154 Caucasian GD patients and 118 Caucasian controls showed an association between the CC genotype and GD but with a low RR of 1.6 (252). TDT analysis also showed preferential transmission of the C allele of the CD40 Kozak SNP to affected individuals

(252). However, it is possible that other polymorphisms in the CD40 gene or another gene in LD with CD40 is the GD susceptibility gene in this region.

VII. Whole Genome Screening in AITD

Recently, two whole genome scans were reported in AITD, one in 56 multigenerational Caucasian families (193) and another in 123 Japanese sib-pair families (134). In both studies, the linkage was analyzed for three different phenotypes: GD, HT, and AITD (*i.e.*, GD+HT), and in both studies many loci gave low positive LOD scores (LOD scores >1.0 and <2.0), and only a few loci gave significant LOD scores (>2.0).

The Japanese whole genome screen identified two loci giving strong evidence for linkage (*i.e.*, MLS > 2.0). One locus on chromosome 5q31 showed evidence for linkage with AITD with an MLS of 3.14, and a second locus on chromosome 8q24 showed evidence for linkage with both AITD (MLS = 2.31) and HT (MLS = 3.77) (134). The identification of the Tg gene as the susceptibility gene in this locus has been discussed earlier (see Section V.C). The 5q31 locus contains a cytokine gene cluster, and therefore, several positional candidate genes exist in this locus and need to be examined.

Our genome screen in the Caucasian families was recently expanded to include 102 multigenerational families (540 individuals) (228). In the expanded dataset seven loci were identified, four of them loci that were identified in the original dataset of 56 families and replicated in the expanded dataset (228). Three loci on chromosomes 6p, 8q, and 10q showed evidence for linkage with both GD and HT (MLS = 2.0, 3.5, and 4.1, respectively); three loci showed evidence for linkage with GD, *i.e.*, on 7q (MLS = 2.3), 14q (MLS = 2.1), and 20q (MLS = 3.3); and one locus on 12q showed evidence of linkage with HT (MLS = 3.4) (228). Four of the loci (the AITD locus on 6p, the GD loci on 14q and 20q, and the HT locus on 12q) were identified in the original dataset and replicated in the expanded dataset (228). These four loci represent strong candidates for harboring susceptibility genes for AITD.

Two additional whole genome screens, performed in two single large pedigrees, were also recently reported. One study was performed in a large family with multiple members affected with vitiligo and HT (307). The study identified an autoimmunity locus on 1p31–32 and an additional HT locus that was mapped to the same position as the 6p locus identified in our whole genome screen (designated AITD-1) (Table 7). The second study was performed in a Chinese-American family and identified two loci on chromosomes 9 and 11 that were different from loci reported in Caucasian families (257). Another very recent whole genome scan showed evidence for linkage with 5q31 (304) replicating the results of the Japanese study (134).

VIII. Mechanisms by Which Genes Can Induce Thyroid Autoimmunity

A. General principles

In classical monogenic diseases, the genetic defect changes the action of a gene by decreasing its effectiveness [*e.g.*, the

Pendrin gene in Pendred syndrome (58)] or causing overactivity in the gene [*e.g.*, the RET protooncogene in multiple endocrine neoplasia 2 (258, 259)]. However, in multifactorial diseases such as the AITD, the situation is more complex. The genetic defect may cause subtle changes in the function of one or more genes that when combined increase the likelihood of an individual to develop the disease. Therefore, even when a gene causing a common disease is mapped, proving that the polymorphism is also biologically meaningful can be difficult. But only after it is demonstrated that alterations in the functions of this gene are involved in the pathogenesis of the disease can the gene be declared conclusively a susceptibility gene for the disease.

Conclusive evidence for genes that confer susceptibility to complex diseases has been found. The two best examples for genes that have been shown to predispose to complex diseases are the HLA-DR and DQ genes in type 1 DM (for review, see Ref. 260) and the NOD2 gene in Crohn's disease (60, 84). Recently, some functional studies have been published in AITD, although the results are not yet conclusive (261, 262).

B. HLA

The mechanisms by which HLA molecules confer susceptibility to autoimmune diseases are now beginning to be understood. T cells recognize and respond to an antigen by interacting with a complex between an antigenic peptide and an HLA molecule (reviewed in Ref. 263). It is thought that different HLA alleles have different affinities for peptides from autoantigens (*e.g.*, thyroid antigens) that are recognized by T cell receptors on cells that have escaped tolerance (87). Thus, certain alleles may permit the autoantigenic peptide to fit into the antigen binding groove inside the HLA molecule and to be recognized by the T cell receptor, whereas others may not (264). This would determine whether an autoimmune response to that antigen will develop.

Studies on the structure of HLA polymorphisms associated with type 1 DM provided strong evidence in support of this hypothesis. Sequencing of the HLA DQ genes showed that an aspartic residue at position 57 of the DQ β chain played a key role in the genetic susceptibility to type 1 diabetes (260). Individuals who did not have Asp (57) on both of their DR alleles were at high risk for type 1 diabetes (RR > 50) (265). Moreover, it has been shown that an aspartic acid at position 57 on the DQ β chain influences the antigen binding properties of the HLA-DQ $\alpha\beta$ heterodimer (260, 266). Lack of aspartic acid at position 57 on the DQ β chain permitted immunogenic insulin peptides to fit into the antigen binding groove inside the HLA molecule and to be recognized by the T cell receptor (267, 268). In contrast, the presence of aspartic acid at position 57 of the DQ β chain prevented insulin peptides from fitting and hence prevented autoantigen presentation to the T cell receptor (264).

GD is associated with HLA-DR3 (Table 3). We sequenced the DR3 β chain to identify critical amino acids for the susceptibility to GD (112). Analysis of the frequencies of DR3-specific amino acids occupying the peptide binding pockets in GD patients and controls showed that lack of arginine at position 74 of the DR β chain (DR β 74^{Arg}) was significantly

more frequent in the DR3-positive controls than in the DR3-positive patients (112). Although some HLA-DR binding studies have shown higher affinity of HLA-DR3 to TSHR immunodominant peptides than to TSHR nonimmunodominant peptides, this approach has not been applied to the different DR3 subtypes associated with GD. Nevertheless, such data have suggested that certain DR sequences influence the binding and presentation of TSHR peptides (269), and this may provide a mechanism by which DR β 74^{Arg} influences the susceptibility to GD (Fig. 4). These results, if confirmed, may indicate a general principle in HLA-induced susceptibility to autoimmune diseases as seen in type 1 DM.

For thyroid autoantigens to be presented by HLA molecules to T cells, a mechanism of autoantigen presentation must exist within the thyroid gland or the draining lymph nodes of the gland. One potential intrathyroidal mechanism not using professional APC may be through aberrant expression of HLA class II molecules on thyrocytes (270–272). Unlike in normal thyroids, the thyroid epithelial cells from patients with GD and HT have been shown to express HLA class II antigen molecules similar to those normally expressed on APC such as macrophages and dendritic cells (270, 273) (reviewed in Ref. 274). This aberrant expression of HLA class II molecules on thyroid cells may initiate thyroid autoimmunity via direct thyroid autoantigen presentation (270, 275) or a secondary event after cytokine secretion by invading T cells. Consistent with the former possibility was the fact that thyroid cell MHC class II antigen expression could be induced by certain viral infections *in vitro* (276, 277) and that mice constitutively expressing thyroid cell MHC class II antigens developed thyroiditis after immunization with human Tg (125). Furthermore, a murine model of GD has been shown to depend on TSHR antigen presentation on cells expressing MHC class II molecules (278, 279). Coculture of peripheral blood mononuclear cell from GD patients with homologous thyrocytes induced T cell activation (280) as well as interferon- γ production and thyroid cell HLA class II antigen expression (281). Such cytokine secretion may be the common cause of HLA class II antigen expression by thyroid cells in AITD (271, 282; reviewed in Ref. 283).

C. CTLA-4

The CTLA-4 gene polymorphisms have also been studied for their effects on CTLA-4 function. CTLA-4 is an important costimulatory molecule that participates in the presentation of peptides to T cells. APC activate T cells by presenting to the T cell receptor an antigenic peptide bound to an HLA class II protein on the cell surface. However, a second signal is also required for T cell activation, and these costimulatory signals may be provided by the APC themselves or other local cells (137). The costimulatory signals are provided by a variety of proteins (*e.g.*, B7-1, B7-2, CD40) that are expressed on APC and interact with receptors (CD28, CTLA-4, and CD40L) on the surface of CD4⁺ T lymphocytes during antigen presentation (137). Whereas the binding of B7 to CD28 on T cells costimulates T cell activation, the higher affinity binding of B7 to CTLA-4 down-regulates T cell activation and induces tolerance (Fig. 3). The suppressive effects of CTLA-4 on T cell activation have raised the possibility that the

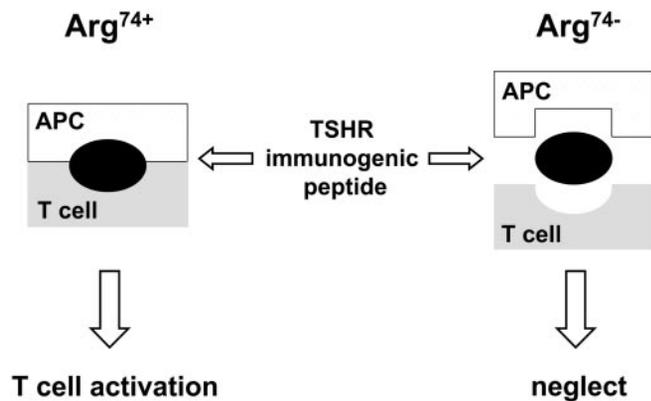


FIG. 4. Possible mechanism of induction of GD by specific HLA-DR sequences. HLA-DR molecules containing Arginine at position 74 of the DR β 1 chain (DR β 74^{Arg}) form a peptide binding pocket that enables presentation of TSHR immunogenic peptides which stimulate TSHR-specific T cells. In contrast, HLA-DR molecules lacking the Arginine at position 74 of the DR β 1 chain cannot fit the TSHR immunogenic peptides, and therefore TSHR-specific T cells are not stimulated.

CTLA-4 polymorphisms associated with AITD decreased its expression and/or function, thereby promoting the development of autoimmunity (151).

As discussed earlier, two CTLA-4 polymorphisms have been shown to be associated with AITD, a 3' UTR microsatellite and an A/G polymorphism in the leader sequence of the gene. One recent study (261) examined the effects of the A and G alleles of the CTLA-4 A/G₄₉ SNP on the inhibitory function of CTLA-4. The authors showed that blocking of CTLA-4 on T cells isolated from individuals with the G allele had less effect on reducing the inhibitory function of CTLA-4 than blocking CTLA-4 on T cells isolated from individuals with the A allele (261). This could imply that the A and G alleles of the CTLA-4 leader sequence influenced its function and/or expression. We have examined the effects of the CTLA-4 A/G₄₉ SNP using an *in vitro* assay by transfecting T cell lines lacking CTLA-4 with CTLA-4 cDNA having the A or the G allele. When T cells were transfected with CTLA-4 cDNA carrying the G or A allele, there was no difference in the expression and inhibitory function of CTLA-4 (262). We concluded that the A and G alleles of the CTLA-4 A/G₄₉ SNP did not directly influence its function. Other polymorphisms must be responsible for the association of CTLA-4 with AITD. However, our results did not necessarily contradict the published data because they used a dissimilar study design. Kouki *et al.* (261) studied the differences in the function of T cells taken from individuals with the A and the G alleles. However, the differences they found could be due to another CTLA-4 polymorphism that was in LD with the A/G₄₉ polymorphism. Indeed, preliminary data in myasthenia gravis showed that the AT microsatellite at the 3' UTR of the CTLA-4 gene influenced the half-life of the CTLA-4 mRNA (284, 285). This could provide an attractive explanation for the association between the short alleles of the AT microsatellite and AITD, as well as other autoimmune diseases.

D. Hypothetical mechanisms by which Tg could induce susceptibility to AITD

As mentioned above (see Section V.C), the Tg gene is linked and associated with AITD (134, 214). Therefore, Tg may be a susceptibility gene for AITD. To demonstrate that Tg is indeed the AITD susceptibility gene on chromosome 8q24, we have sequenced the gene in patients and controls and identified the sequence variants that are associated with AITD. The Tg gene may predispose to AITD in a number of ways, for example: 1) sequence changes in Tg may change its antigenicity, making certain Tg peptides more immunogenic; 2) sequence changes in Tg may change its interaction with HLA molecules; and 3) sequence changes in or near the Tg gene may alter its expression, and this may lead to reduced immune tolerance to the Tg molecule. In addition, alterations in Tg could possibly explain interactions between genetic and environmental factors in the etiology of AITD, because Tg is iodinated to form thyroid hormones, and dietary iodine may influence the development of AITD (286–288). Indeed, as noted above, the Tg hormonogenic sites were shown to contain the autoepitopes in EAT, although the role of iodine is still controversial in experimental thyroiditis (206, 207).

IX. Conclusions and Future Directions

The AITD are complex diseases believed to be caused by the combined effects of multiple susceptibility genes and environmental triggers (Fig. 5). There are sufficient epidemiological data to support an important genetic contribution to the development of AITD, and in the past few years several loci and genes have shown evidence for linkage and/or association with AITD. Thus, the genetic susceptibility to AITD seems to involve several genes with varying effects. With the completion of the Human Genome Project and the establishment of large SNP databases, the identification of additional AITD susceptibility genes will become more feasible. One approach that was proposed is to perform whole genome association studies using several thousand SNPs by pooling DNA samples (72).

The AITD loci identified so far show that some putative

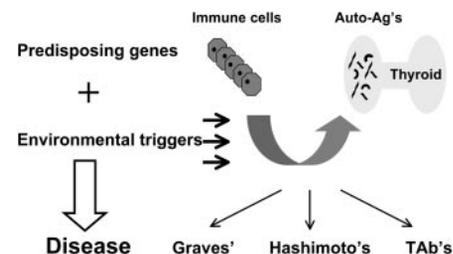


FIG. 5. A proposed mechanism for the development of AITD. Individuals who are born with susceptibility genes are predisposed to develop AITD. The AITD susceptibility genes include sequence variants in immune-regulatory genes or in thyroid-specific genes (*e.g.*, Tg). A genetically susceptible individual can then encounter an environmental triggering factor [*e.g.*, infection, dietary factors (iodine), stress, and pregnancy]. This encounter in a genetically susceptible individual leads to the development of the clinical phenotype of GD, HT, or thyroid antibodies alone. The final clinical phenotype is determined by both the susceptibility genes and the environmental factors.

AITD susceptibility genes may be immune modifying genes that increase the susceptibility to autoimmunity in general (e.g., HLA, CTLA-4), whereas others may be specific to AITD (e.g., TSHR, Tg). The next step in investigating the role of these genes in the development of AITD is by functional studies and genotype-phenotype correlations. Preliminary functional studies have been performed for HLA (112, 269) and CTLA-4 (151, 261, 262). More functional studies are needed for these and other genes that have shown association with AITD.

It is most likely that the susceptibility loci for AITD interact and that their interactions may influence disease phenotype and severity (193). The molecular basis for the interactions between susceptibility genes in complex diseases is unknown. These interactions could represent the cumulative effect of increased statistical risk, or alternatively, there may be molecular interactions between the susceptibility genes or their products, which ultimately determine disease phenotype. Another unresolved question is how do environmental factors interact with susceptibility genes to modify the risk for disease, as well as the disease phenotype. We are slowly progressing toward identification of the AITD susceptibility genes, and once they are identified we will begin to understand the underlying molecular mechanisms by which they induce thyroid autoimmunity.

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References

1. **Davies TF** 2000 Graves' diseases: pathogenesis. In: Braverman LE, Utiger RD, eds. *Werner and Ingbar's the thyroid: a fundamental and clinical text*. Philadelphia: Lippincott Williams & Wilkins; 518–530
2. **Weetman AP** 1996 Chronic autoimmune thyroiditis. In: Braverman LE, Utiger RD, eds. *Werner and Ingbar's the thyroid*. Philadelphia: Lippincott-Raven; 738–748
3. **Stagnaro-Green A** 2002 Clinical review 152: postpartum thyroiditis. *J Clin Endocrinol Metab* 87:4042–4047
4. **Roti E, Uberti E** 2002 Post-partum thyroiditis—a clinical update. *Eur J Endocrinol* 146:275–279
5. **Muller AF, Drexhage HA, Berghout A** 2001 Postpartum thyroiditis and autoimmune thyroiditis in women of childbearing age: recent insights and consequences for antenatal and postnatal care. *Endocr Rev* 22:605–630
6. **Martino E, Bartalena L, Bogazzi F, Braverman LE** 2001 The effects of amiodarone on the thyroid. *Endocr Rev* 22:240–254
7. **Marazuela M, Garcia-Buey L, Gonzalez-Fernandez B, Garcia-Monzon C, Arranz A, Borque MJ, Moreno-Otero R** 1996 Thyroid autoimmune disorders in patients with chronic hepatitis C before and during interferon- α therapy. *Clin Endocrinol (Oxf)* 44:635–642
8. **Dumoulin FL, Leifeld L, Sauerbruch T, Spengler U** 1999 Autoimmunity induced by interferon- α therapy for chronic viral hepatitis. *Biomed Pharmacother* 53:242–254
9. **Obermayer-Straub P, Manns MP** 1998 Autoimmune polyglandular syndromes. *Baillieres Clin Gastroenterol* 12:293–315
10. **Betterle C, Volpato M, Greggio AN, Presotto F** 1996 Type 2 polyglandular autoimmune disease (Schmidt's syndrome). *J Pediatr Endocrinol Metab* 9(Suppl 1):113–123
11. **Hall R, Dingle PR, Roberts DF** 1972 Thyroid antibodies: a study of first degree relatives. *Clin Genet* 3:319–324
12. **Brix TH, Kyvik KO, Hegedus L** 1998 What is the evidence of genetic factors in the etiology of Graves' disease? A brief review. *Thyroid* 8:727–734
13. **Tomer Y, Barbesino G, Greenberg DA, Davies TF** 1997 The immunogenetics of autoimmune diabetes and autoimmune thyroid disease. *Trends Endocrinol Metab* 8:63–70
14. **Hollowell JG, Staehling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA, Braverman LE** 2002 Serum TSH, T(4), and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). *J Clin Endocrinol Metab* 87:489–499
15. **Canaris GJ, Manowitz NR, Mayor G, Ridgway EC** 2000 The Colorado thyroid disease prevalence study. *Arch Intern Med* 160:526–534
16. **Tunbridge WMG, Evered DC, Hall R, Appleton D, Brewis M, Clark F, Evans JG, Young E, Bird T, Smith PA** 1977 The spectrum of thyroid disease in a community: the Whickham survey. *Clin Endocrinol (Oxf)* 7:481–493
17. **Phillips DI, Barker DJ, Rees Smith B, Didcote S, Morgan D** 1985 The geographical distribution of thyrotoxicosis in England according to the presence or absence of TSH-receptor antibodies. *Clin Endocrinol (Oxf)* 23:283–287
18. **Brownlie BE, Welsh JD** 1990 The epidemiology of thyrotoxicosis in New Zealand: incidence and geographical distribution in north Canterbury, 1983–1985. *Clin Endocrinol (Oxf)* 33:249–259
19. **Berglund J, Christensen SB, Hallengren B** 1990 Total and age-specific incidence of Graves' thyrotoxicosis, toxic nodular goitre and solitary toxic adenoma in Malmö 1970–74. *J Intern Med* 227:137–141
20. **Laurberg P, Pedersen KM, Vestergaard H, Sigurdsson G** 1991 High incidence of multinodular toxic goitre in the elderly population in a low iodine intake area vs. high incidence of Graves' disease in the young in a high iodine intake area: comparative surveys of thyrotoxicosis epidemiology in East-Jutland Denmark and Iceland. *J Intern Med* 229:415–420
21. **Bulow Pedersen I, Knudsen N, Jorgensen T, Perrild H, Ovesen L, Laurberg P** 2002 Large differences in incidences of overt hyper- and hypothyroidism associated with a small difference in iodine intake: a prospective comparative register-based population survey. *J Clin Endocrinol Metab* 87:4462–4469
22. **Haraldsson A, Gudmundsson ST, Larusson G, Sigurdsson G** 1985 Thyrotoxicosis in Iceland 1980–1982. An epidemiological survey. *Acta Med Scand* 217:253–258
23. **Furszyfer J, Kurland LT, McConahey WM, Elveback LR** 1970 Graves' disease in Olmsted County, Minnesota, 1935 through 1967. *Mayo Clin Proc* 45:636–644
24. **Mogensen EF, Green A** 1980 The epidemiology of thyrotoxicosis in Denmark. Incidence and geographical variation in the Funen region 1972–1974. *Acta Med Scand* 208:183–186
25. **Gordin A, Heinonen OP, Saarinen P, Lamberg BA** 1972 Serum thyrotrophin in symptomless autoimmune thyroiditis. *Lancet* 1:551–554
26. **Okamura K, Ueda K, Sone H, Ikenoue H, Hasuo Y, Sato K, Yoshinari M, Fujishima M** 1989 A sensitive thyroid stimulating hormone assay for screening of thyroid functional disorder in elderly Japanese. *J Am Geriatr Soc* 37:317–322
27. **Jacobson DL, Gange SJ, Rose NR, Graham NM** 1997 Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol* 84:223–243
28. **Sawin CT, Castelli WP, Hershman JM, McNamara P, Bacharach P** 1985 The aging thyroid. Thyroid deficiency in the Framingham Study. *Arch Intern Med* 145:1386–1388
29. **Furszyfer J, Kurland LT, McConahey WM, Woolner LB, Elveback LR** 1972 Epidemiologic aspects of Hashimoto's thyroiditis and

- Graves' disease in Rochester, Minnesota (1935–1967), with special reference to temporal trends. *Metabolism* 21:197–204
30. **Berglund J, Ericsson UB, Hallgren B** 1996 Increased incidence of thyrotoxicosis in Malmo during the years 1988–1990 as compared to the years 1970–1974. *J Intern Med* 239:57–62
 31. **Davies TF, Amino N** 1993 A new classification for human autoimmune thyroid disease. *Thyroid* 3:331–333
 32. **Bartels ED** 1941 Twin examinations: heredity in Graves' disease. Copenhagen: Munksgaard; 32–36
 33. **Martin L** 1945 The heredity and familial aspects of exophthalmic goitre and nodular goitre. *Q J Med* 14:207–219
 34. **Hall R, Stanbury JB** 1967 Familial studies of autoimmune thyroiditis. *Clin Exp Immunol* 2:719–725
 35. **Tamai H, Ohsako N, Takeno K, Fukino O, Takahashi H, Kuma K, Kumagai LF, Nagataki S** 1980 Changes in thyroid function in euthyroid subjects with family history of Graves' disease; a follow-up study of 69 patients. *J Clin Endocrinol Metab* 51:1123–1128
 36. **Chopra IJ, Solomon DH, Chopra U, Yodhishara E, Tersaki PL, Smith F** 1977 Abnormalities in thyroid function in relatives of patients with Graves' disease and Hashimoto's thyroiditis: lack of correlation with inheritance of HLA-B8. *J Clin Endocrinol Metab* 45:45–54
 37. **Tamai H, Kumagai LF, Nagataki S** 1986 Immunogenetics of Graves' disease. In: McGregor AM, ed. *Immunology of endocrine diseases*. Lancaster, UK: MTP Press; 123–141
 38. **Carey C, Skosey C, Pinnamaneni KM, Barsano CP, DeGroot LJ** 1980 Thyroid abnormalities in children of parents who have Graves' disease: possible pre-Graves' disease. *Metabolism* 29:369–376
 39. **Volpe R** 1985 Autoimmune thyroid disease. In: Volpe R, ed. *Autoimmunity and endocrine disease*. New York: Marcel Dekker; 109–285
 40. **Burek CL, Hoffman WH, Rose NR** 1982 The presence of thyroid autoantibodies in children and adolescents with AITD and in their siblings and parents. *Clin Immunol Immunopathol* 25:395–404
 41. **Villanueva RB, Inzerillo AM, Tomer Y, Barbesino G, Meltzer M, Concepcion ES, Greenberg DA, Maclaren N, Sun ZS, Zhang DM, Tucci S, Davies TF** 2000 Limited genetic susceptibility to severe Graves' ophthalmopathy: no role for CTLA-4 and evidence for an environmental etiology. *Thyroid* 10:791–798
 42. **Tomer Y, Davies TF** 1993 Infection, thyroid disease and autoimmunity. *Endocr Rev* 14:107–120
 43. **Williams D** 1996 Thyroid cancer and the Chernobyl accident. *J Clin Endocrinol Metab* 81:6–8
 44. **Risch N** 1990 Linkage strategies for genetically complex traits. II. The power of affected relative pairs. *Am J Hum Genet* 46:229–241
 45. **Villanueva R, Greenberg DA, Davies TF, Tomer Y** 2003 Sibling recurrence risk in autoimmune thyroid disease. *Thyroid* 13:761–764
 46. **Allahabadia A, Gough SC** 1999 The different approaches to the genetic analysis of autoimmune thyroid disease. *J Endocrinol* 163:7–13
 47. **Pauls DL, Zakarija M, McKenzie JM, Egeland JA** 1993 Complex segregation analysis of antibodies to thyroid peroxidase in Old Order Amish families. *Am J Med Genet* 47:375–379
 48. **Phillips D, McLachlan S, Stephenson A, Roberts D, Moffitt S, McDonald D, Ad'Hiiah A, Stratton A, Young E, Clark F** 1990 Autosomal dominant transmission of autoantibodies to thyroglobulin and thyroid peroxidase. *J Clin Endocrinol Metab* 70:742–746
 49. **Phillips DIW, Prentice L, McLachlan SM, Upadhyaya M, Lunt PW, Rees Smith B** 1991 Autosomal dominant inheritance of the tendency to develop thyroid autoantibodies. *Exp Clin Endocrinol* 97:170–172
 50. **Jaume JC, Guo J, Pauls DL, Zakarija M, McKenzie JM, Egeland JA, Burek CL, Rose NR, Hoffman WH, Rapoport B, McLachlan SM** 1999 Evidence for genetic transmission of thyroid peroxidase autoantibody epitopic "fingerprints." *J Clin Endocrinol Metab* 84:1424–1431
 51. **Hall JG** 1996 Twinning: mechanisms and genetic implications. *Curr Opin Genet Dev* 6:343–347
 52. **Monteiro J, Hingorani R, Choi I-H, Silver J, Pergolizzi R, Gergersen PK** 1995 Oligoclonality in the human CD8+ T cell repertoire in normal subjects and twins: implications for studies of infectious and autoimmune diseases. *Mol Med* 1:614–624
 53. **Brix TH, Christensen K, Holm NV, Harvald B, Hegedus L** 1998 A population-based study of Graves' diseases in Danish twins. *Clin Endocrinol (Oxf)* 48:397–400
 54. **Brix TH, Kyvik KO, Christensen K, Hegedus L** 2001 Evidence for a major role of heredity in Graves' disease: a population-based study of two Danish twin cohorts. *J Clin Endocrinol Metab* 86:930–934
 55. **Ringold DA, Nicoloff JT, Kesler M, Davis H, Hamilton A, Mack T** 2002 Further evidence for a strong genetic influence on the development of autoimmune thyroid disease: the California twin study. *Thyroid* 12:647–653
 56. **Brix TH, Kyvik KO, Hegedus L** 2000 A population-based study of chronic autoimmune hypothyroidism in Danish twins. *J Clin Endocrinol Metab* 85:536–539
 57. **Phillips DI, Osmond C, Baird J, Huckle A, Rees-Smith B** 2002 Is birthweight associated with thyroid autoimmunity? A study in twins. *Thyroid* 12:377–380
 58. **Everett LA, Glaser B, Beck JC, Idol JR, Buchs A, Heyman M, Adawi F, Hazani E, Nassir E, Baxevanis AD, Sheffield VC, Green ED** 1997 Pendred syndrome is caused by mutations in a putative sulphate transporter gene (PDS). *Nat Genet* 17:411–422
 59. **Davies JL, Kawauchi Y, Bennet ST, Copeman JB, Cordell HJ, Pritchard LE, Reed PW, Gough SCL, Jenkins SC, Palmer SM, Balfour KM, Rowe BR, Farrall M, Barnett AH, Baln SC, Todd JA** 1994 A genome-wide search for human type 1 diabetes susceptibility genes. *Nature* 371:130–136
 60. **Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, Binder V, Finkel Y, Cortot A, Modigliani R, Laurent-Puig P, Gower-Rousseau C, Macry J, Colombel J-F, Sahbatou M, Thomas G** 2001 Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 411:599–603
 61. **Greenberg DA** 1993 Linkage analysis of "necessary" loci vs. "susceptibility" loci. *Am J Hum Genet* 52:135–143
 62. **Ott J** 1996 Analysis of human genetic linkage. Baltimore: Johns Hopkins University Press
 63. **Greenberg DA, Abreu PC** 2001 Determining trait locus position from multipoint analysis: accuracy and power of three different statistics. *Genet Epidemiol* 21:299–314
 64. **Terwilliger JD, Zollner S, Laan M, Paabo S** 1998 Mapping genes through the use of linkage disequilibrium generated by genetic drift: 'drift mapping' in small populations with no demographic expansion. *Hum Hered* 48:138–154
 65. **Hodge SE, Anderson CE, Neiswanger K, Sparkes RS, Rimoin DL** 1983 The search for heterogeneity in insulin dependent diabetes mellitus (IDDM): linkage studies, two-locus models, and genetic heterogeneity. *Am J Hum Genet* 35:1139–1155
 66. **Lander E, Kruglyak L** 1995 Genetic dissection of complex traits: guidelines for interpreting and reporting linkage results. *Nat Genet* 11:241–247
 67. **Pal DK, Greenberg DA** 2002 Evaluating genetic heterogeneity in complex disorders. *Hum Hered* 53:216–226
 68. **Ott J** 1977 Counting methods (EM algorithm) in human pedigree analysis: linkage and segregation analysis. *Ann Hum Genet* 40:443–454
 69. **Risch N, Baron M** 1982 X-linkage and genetic heterogeneity in bipolar-related major affective illness: reanalysis of linkage data. *Ann Hum Genet* 46:153–166
 70. **Kruglyak L, Daly MJ, Reeve-Daly MP, Lander ES** 1996 Parametric and nonparametric linkage analysis: a unified multipoint approach. *Am J Hum Genet* 58:1347–1363
 71. **Risch N, Merikangas K** 1996 The future of genetic studies of complex human diseases. *Science* 273:1516–1517
 72. **Sham P, Bader JS, Craig I, O'Donovan M, Owen M** 2002 DNA pooling: a tool for large-scale association studies. *Nat Rev Genet* 3:862–871
 73. **Woolf B** 1955 On estimating the relation between blood group and disease. *Ann Hum Genet* 19:251–253
 74. **Hodge SE** 1994 What association analysis can and cannot tell us about the genetics of complex disease. *Am J Med Genet* 54:318–323
 75. **Spielman RS, McGinnis RE, Ewens WJ** 1993 Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus. *Am J Hum Genet* 52:506–516

76. Falk CT, Rubinstein P 1987 Haplotype relative risks: an easy and reliable way to construct a proper control sample for risk calculations. *Ann Hum Genet* 51:227–233
77. Schaid DJ, Sommer SS 1994 Comparison of statistics for candidate-gene association studies using cases and parents. *Am J Hum Genet* 55:402–409
78. Greenberg DA, Berger B 1994 Using LOD score differences to determine mode of inheritance: simple, robust method even in the presence of heterogeneity and reduced penetrance. *Am J Hum Genet* 55:834–840
79. Tomer Y, Barbesino G, Greenberg DA, Concepcion ES, Davies TF 1998 A new Graves' disease-susceptibility locus maps to chromosome 20q11.2. *Am J Hum Genet* 63:1749–1756
80. Froguel P, Zouali H, Vionnet N, Velho G, Vaxillaire M, Pharm D, Sun F, Lesage S, Stoffel M, Takeda J, Passa P, Permutt MA, Beckmann J, Bell GI, Cohen D 1993 Familial hyperglycemia due to mutations in glucokinase. Definition of a subtype of diabetes mellitus. *N Engl J Med* 328:697–702
81. Weber JL 1990 Human DNA polymorphisms based on length variations in simple-sequence tandem repeats. In: Tilghman S, Davies K, eds. *Genome analysis, Vol 1. Genetic and physical mapping*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 159–181
82. Brookes AJ 1999 The essence of SNPs. *Gene* 234:177–186
83. Laan M, Paabo S 1998 Mapping genes by drift-generated linkage disequilibrium. *Am J Hum Genet* 63:654–656
84. Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, Achkar J-P, Brant SR, Bayless TM, Kirschner BS, Hanauer SB, Nuñez G, Cho JH 2001 A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 411:603–606
85. Morahan G, Huang D, Ymer SI, Cancilla MR, Stephen K, Dabadghao P, Werther G, Tait BD, Harrison LC, Colman PG 2001 Linkage disequilibrium of a type 1 diabetes susceptibility locus with a regulatory IL12B allele. *Nat Genet* 27:218–221
86. Campbell RD, Trowsdale J 1993 Map of the human MHC. *Immunol Today* 14:349–352
87. Nelson JL, Hansen JA 1990 Autoimmune disease and HLA. *CRC Crit Rev Immunol* 10:307–328
88. Bech K, Lumholtz B, Nerup J, Thomsen M, Platz P, Ryder LP, Svejgaard A, Siersbaek-Nielsen K, Hansen JM, Larse JH 1977 HLA antigens in Graves' disease. *Acta Endocrinol (Copenh)* 86:510–516
89. Farid NR, Stone E, Johnson G 1980 Graves' disease and HLA: clinical and epidemiologic associations. *Clin Endocrinol (Oxf)* 13:535–544
90. Farid NR 1981 Graves' disease. In: Farid NR, ed. *HLA in endocrine and metabolic disorders*. New York: Academic Press; 85–143
91. Farid NR, Sampson L, Noel EP, Barnard JM, Mandeville R, Larsen B, Marshall WH, Carter ND 1979 A study of human D locus related antigens in Graves' disease. *J Clin Invest* 63:108–113
92. Mangklabruks A, Cox N, DeGroot LJ 1991 Genetic factors in autoimmune thyroid disease analyzed by restriction fragment length polymorphisms of candidate genes. *J Clin Endocrinol Metab* 73:236–244
93. Volpe R 1990 Immunology of human thyroid disease. In: Volpe R, ed. *Autoimmunity in endocrine disease*. Boca Raton, FL: CRC Press; 73
94. Heward JM, Allahabadia A, Daykin J, Carr-Smith J, Daly A, Armitage M, Dodson PM, Sheppard MC, Barnett AH, Franklyn JA, Gough SC 1998 Linkage disequilibrium between the human leukocyte antigen class II region of the major histocompatibility complex and Graves' disease: replication using a population case control and family-based study. *J Clin Endocrinol Metab* 83:3394–3397
95. Kawa A, Nakamura S, Nakazawa M, Sakaguchi S, Kawabata T, Maeda Y, Kanehisa T 1977 HLA-BW35 and B5 in Japanese patients with Graves' disease. *Acta Endocrinol (Copenh)* 86:754–757
96. Inoue D, Sato K, Enomoto T, Sugawa H, Maeda M, Inoko H, Tsuji K, Mori T, Imura H 1992 Correlation of HLA types and clinical findings in Japanese patients with hyperthyroid Graves' disease: evidence indicating the existence of four subpopulations. *Clin Endocrinol (Oxf)* 36:75–82
97. Onuma H, Ota M, Sugeno A, Inoko H 1994 Association of HLA-DPB1*0501 with early-onset Graves' disease in Japanese. *Hum Immunol* 39:195–201
98. Katsuren E, Awata T, Matsumoto C, Yamamoto K 1994 HLA class II alleles in Japanese patients with Graves' disease: weak associations of HLA-DR and -DQ. *Endocr J* 41:599–603
99. Ohtsuka K, Nakamura Y 1998 Human leukocyte antigens associated with hyperthyroid Graves ophthalmology in Japanese patients. *Am J Ophthalmol* 126:805–810
100. Chan SH, Yeo PP, Lui KF, Wee GB, Woo KT, Lim P, Cheah JS 1978 HLA and thyrotoxicosis (Graves' disease) in Chinese. *Tissue Antigens* 12:109–114
101. Cavan DA, Penny MA, Jacobs KH, Kelly MA, Jenkins D, Mijovic C, Chow C, Cockram CS, Hawkins BR, Barnett AH 1994 The HLA association with Graves' disease is sex-specific in Hong Kong Chinese subjects. *Clin Endocrinol (Oxf)* 40:63–66
102. Chan SH, Lin YN, Wee GB, Ren EC, Lui KF, Cheah JS 1993 Human leukocyte antigen DNA typing in Singaporean Chinese patients with Graves' disease. *Ann Acad Med Singapore* 22:576–579
103. Tan S, Chan S, Lee B, Wee G, Wong H 1988 HLA association in Singapore children with Grave's disease. *Metabolism* 37:518–519
104. Yeo PP, Chan SH, Thai AC, Ng WY, Lui KF, Wee GB, Tan SH, Lee BW, Wong HB, Cheah JS 1989 HLA Bw46 and DR9 associations in Graves' disease of Chinese patients are age- and sex-related. *Tissue Antigens* 34:179–184
105. Glazier AM, Nadeau JH, Aitman TJ 2002 Finding genes that underlie complex traits. *Science* 298:2345–2349
106. Maciel LM, Rodrigues SS, Dibbern RS, Navarro PA, Donadi EA 2001 Association of the HLA-DRB1*0301 and HLA-DQA1*0501 alleles with Graves' disease in a population representing the gene contribution from several ethnic backgrounds. *Thyroid* 11:31–35
107. Barlow ABT, Wheatcroft N, Watson P, Weetman AP 1996 Association of HLA-DQA1*0501 with Graves' disease in English Caucasian men and women. *Clin Endocrinol (Oxf)* 44:73–77
108. Yanagawa T, Mangklabruks A, Chang YB, Okamoto Y, Fisfalen M-E, Curran PG, DeGroot LJ 1993 Human histocompatibility leukocyte antigen-DQA1*0501 allele associated with genetic susceptibility to Graves' disease in a Caucasian population. *J Clin Endocrinol Metab* 76:1569–1574
109. Marga M, Denisova A, Sochnev A, Pirags V, Farid NR 2001 Two HLA DRB 1 alleles confer independent genetic susceptibility to Graves disease: relevance of cross-population studies. *Am J Med Genet* 102:188–191
110. Zamani M, Spaepen M, Bex M, Bouillon R, Cassiman JJ 2000 Primary role of the HLA class II DRB1*0301 allele in Graves disease. *Am J Med Genet* 95:432–437
111. Marsh SG, Bodmer JG, Albert ED, Bodmer WF, Bontrop RE, Dupont B, Erlich HA, Hansen JA, Mach B, Mayr WR, Parham P, Petersdorf EW, Sasazuki T, Schreuder GM, Strominger JL, Svejgaard A, Terasaki PI 2001 Nomenclature for factors of the HLA system, 2000. *Hum Immunol* 62:419–468
112. Ban Y, Davies TF, Greenberg DA, Concepcion ES, Tomer Y, Potential role of HLA-DR-74 arginine in the genetic susceptibility to Graves' disease. Program of the 84th Annual Meeting of The Endocrine Society, San Francisco, CA, 2002, p 95 (Abstract OR22-5)
113. Segni M, Pani MA, Pasquino AM, Badenhop K 2002 Familial clustering of juvenile thyroid autoimmunity: higher risk is conferred by human leukocyte antigen DR3-DQ2 and thyroid peroxidase antibody status in fathers. *J Clin Endocrinol Metab* 87:3779–3782
114. Schleusener H, Schwander J, Fischer C, Holle R, Holl G, Badenhop K, Hensen J, Finke R, Bogner U, Mayr WR 1989 Prospective multicentre study on the prediction of relapse after antithyroid drug treatment in patients with Graves' disease. *Acta Endocrinol (Copenh)* 120:689–701
115. Dahlberg PA, Holmlund G, Karlsson FA, Safwenberg J 1981 HLA-A, -B, -C and -DR antigens in patients with Graves' disease and their correlation with signs and clinical course. *Acta Endocrinol (Copenh)* 97:42–47
116. McKenna R, Kearns M, Sugrue D, Drury MI, McCarthy CF 1982 HLA and hyperthyroidism in Ireland. *Tissue Antigens* 19:97–99
117. Allannic H, Fauchet R, Lorc Y, Gueguen M, Le Guerrier AM, Genetet B 1983 A prospective study of the relationship between

- relapse of hyperthyroid Graves' disease after antithyroid drugs and HLA haplotype. *J Clin Endocrinol Metab* 57:719–722
118. **Kendall-Taylor P, Stephenson A, Stratton A, Papiha SS, Perros P, Roberts DF** 1988 Differentiation of autoimmune ophthalmopathy from Graves' hyperthyroidism by analysis of genetic markers. *Clin Endocrinol (Oxf)* 28:601–610
 119. **Rapport B, Alsabeh R, Aftergood D, McLachlan SM** 2000 Elephantiasic pretibial myxedema: insight into and a hypothesis regarding the pathogenesis of the extrathyroidal manifestations of Graves' disease. *Thyroid* 10:685–692
 120. **Irvine WJ, Gray RS, Morris PJ, Ting A** 1978 HLA in primary atrophic hypothyroidism and Hashimoto goitre. *J Clin Lab Immunol* 3:193–195
 121. **Farid NR, Sampson L, Moens H, Barnard JM** 1981 The association of goitrous autoimmune thyroiditis with HLA-DR5. *Tissue Antigens* 17:265–268
 122. **Moens H, Farid NR, Sampson L, Noel EP, Barnard JM** 1978 Hashimoto's thyroiditis is associated with HLA-DRw3. *N Engl J Med* 299:133–134
 123. **Tandon N, Zhang L, Weetman AP** 1991 HLA associations with Hashimoto's thyroiditis. *Clin Endocrinol (Oxf)* 34:383–386
 124. **Ban Y, Davies TF, Greenberg DA, Concepcion ES, Tomer Y** 2002 The influence of human leucocyte antigen (HLA) genes on autoimmune thyroid disease (AITD): results of studies in HLA-DR3 positive AITD families. *Clin Endocrinol (Oxf)* 57:81–88
 125. **Kong YC, Lomo LC, Motte RW, Giraldo AA, Baisch J, Strauss G, Hammerling GJ, David CS** 1996 HLA-DRB1 polymorphism determines susceptibility to autoimmune thyroiditis in transgenic mice: definitive association with HLA-DRB1*0301 (DR3) gene. *J Exp Med* 184:1167–1172
 126. **Wu Z, Stephens HAF, Sachs JA, Biro PA, Cutbush S, Magzoub MM, Becker C, Schwartz G, Botazzo GF** 1994 Molecular analysis of HLA-DQ and -DP genes in caucasoid patients with Hashimoto's thyroiditis. *Tissue Antigens* 43:116–119
 127. **Badenhoop K, Schwartz G, Walfish PG, Drummond V, Usadel KH, Bottazzo GF** 1990 Susceptibility to thyroid autoimmune disease: molecular analysis of HLA-D region genes identifies new markers for goitrous Hashimoto's thyroiditis. *J Clin Endocrinol Metab* 71:1131–1137
 128. **Honda K, Tamai H, Morita T, Kuma K, Nishimura Y, Sasazuki T** 1989 Hashimoto's thyroiditis and HLA in Japanese. *J Clin Endocrinol Metab* 69:1268–1273
 129. **Hawkins BR, Lam KSL, Ma JTC, Wang C, Yeung RTT** 1987 Strong association between HLA-DRw9 and Hashimoto's thyroiditis in Southern Chinese. *Acta Endocrinol (Copenh)* 114:543–546
 130. **Bode HH, Dorf ME, Forbes AP** 1973 Familial lymphocytic thyroiditis: analysis of linkage with histocompatibility and blood group. *J Clin Endocrinol Metab* 37:692–697
 131. **Roman SH, Greenberg DA, Rubinstein P, Wallenstein S, Davies TF** 1992 Genetics of autoimmune thyroid disease: lack of evidence for linkage to HLA within families. *J Clin Endocrinol Metab* 74:496–503
 132. **Hawkins BR, Ma JT, Lam KS, Wang CC, Yeung RT** 1985 Analysis of linkage between HLA haplotype and susceptibility to Graves' disease in multiple-case Chinese families in Hong Kong. *Acta Endocrinol (Copenh)* 110:66–69
 133. **Barbesino G, Tomer Y, Concepcion ES, Davies TF, Greenberg DA** 1998 Linkage analysis of candidate genes in autoimmune thyroid disease. I. Selected immunoregulatory genes. *J Clin Endocrinol Metab* 83:1580–1584
 134. **Sakai K, Shirasawa S, Ishikawa N, Ito K, Tamai H, Kuma K, Akamizu T, Tanimura M, Furugaki K, Yamamoto K, Sasazuki T** 2001 Identification of susceptibility loci for autoimmune thyroid disease to 5q31–q33 and Hashimoto's thyroiditis to 8q23–q24 by multipoint affected sib-pair linkage analysis in Japanese. *Hum Mol Genet* 10:1379–1386
 135. **Vaidya B, Imrie H, Perros P, Young ET, Kelly WF, Carr D, Large DM, Toft AD, McCarthy MI, Kendall-Taylor P, Pearce SH** 1999 The cytotoxic T lymphocyte antigen-4 is a major Graves' disease locus. *Hum Mol Genet* 8:1195–1199
 136. **Shields DC, Ratanachaiyavong S, McGregor AM, Collins A, Morton NE** 1994 Combined segregation and linkage analysis of Graves' disease with a thyroid autoantibody diathesis. *Am J Hum Genet* 55:540–554
 137. **Reiser H, Stadecker MJ** 1996 Costimulatory B7 molecules in the pathogenesis of infectious and autoimmune diseases. *N Engl J Med* 335:1369–1377
 138. **Yanagawa T, Hidaka Y, Guimaraes V, Soliman M, DeGroot LJ** 1995 CTLA-4 gene polymorphism associated with Graves' disease in a Caucasian population. *J Clin Endocrinol Metab* 80:41–45
 139. **Nistico L, Buzzetti R, Pritchard LE, Van der Auwera B, Giovannini C, Bosi E, Larrad MT, Rios MS, Chow CC, Cockram CS, Jacobs K, Mijovic C, Bain SC, Barnett AH, Vandewalle CL, Schuit F, Gorus FK, Tosi R, Pozzilli P, Todd JA** 1996 The CTLA-4 gene region of chromosome 2q33 is linked to, and associated with, type 1 diabetes. *Hum Mol Genet* 5:1075–1080
 140. **Donner H, Rau H, Walfish PG, Braun J, Siegmund T, Finke R, Herwig J, Usadel KH, Badenhoop K** 1997 CTLA4 alanine-17 confers genetic susceptibility to Graves' disease and to type 1 diabetes mellitus. *J Clin Endocrinol Metab* 82:143–146
 141. **Kotsa K, Watson PF, Weetman AP** 1997 A CTLA-4 gene polymorphism is associated with both Graves' disease and autoimmune hypothyroidism. *Clin Endocrinol (Oxf)* 46:551–554
 142. **Kouki T, Gardine CA, Yanagawa T, DeGroot LJ** 2002 Relation of three polymorphisms of the CTLA-4 gene in patients with Graves' disease. *J Endocrinol Invest* 25:208–213
 143. **Nithiyananthan R, Heward JM, Allahabadia A, Franklyn JA, Gough SC** 2002 Polymorphism of the CTLA-4 gene is associated with autoimmune hypothyroidism in the United Kingdom. *Thyroid* 12:3–6
 144. **Braun J, Donner H, Siegmund T, Walfish PG, Usadel KH, Badenhoop K** 1998 CTLA-4 promoter variants in patients with Graves' disease and Hashimoto's thyroiditis. *Tissue Antigens* 51:563–566
 145. **Yanagawa T, Taniyama M, Enomoto S, Gomi K, Maruyama H, Ban Y, Saruta T** 1997 CTLA4 gene polymorphism confers susceptibility to Graves' disease in Japanese. *Thyroid* 7:843–846
 146. **Akamizu T, Sale MM, Rich SS, Hiratani H, Noh JY, Kanamoto N, Saijo M, Miyamoto Y, Saito Y, Nakao K, Bowden DW** 2000 Association of autoimmune thyroid disease with microsatellite markers for the thyrotropin receptor gene and CTLA-4 in Japanese patients. *Thyroid* 10:851–858
 147. **Park YJ, Chung HK, Park DJ, Kim WB, Kim SW, Koh JJ, Cho BY** 2000 Polymorphism in the promoter and exon 1 of the cytotoxic T lymphocyte antigen-4 gene associated with autoimmune thyroid disease in Koreans. *Thyroid* 10:453–459
 148. **Heward JM, Allahabadia A, Armitage M, Hattersley A, Dodson PM, Macleod K, Carr-Smith J, Daykin J, Daly A, Sheppard MC, Holder RL, Barnett AH, Franklyn JA, Gough SC** 1999 The development of Graves' disease and the CTLA-4 gene on chromosome 2q33. *J Clin Endocrinol Metab* 84:2398–2401
 149. **Heward JM, Allahabadia A, Carr-Smith J, Daykin J, Cockram CS, Gordon CBAH, Franklyn JA, Gough SCL** 1998 No evidence for allelic association of human CTLA-4 promoter polymorphism with autoimmune thyroid disease in either population-based case-control or family-based studies. *Clin Endocrinol (Oxf)* 49:331–334
 150. **Tomer Y** 2001 Unraveling the genetic susceptibility to autoimmune thyroid diseases: CTLA-4 takes the stage. *Thyroid* 11:167–169
 151. **Ueda H, Howson JM, Esposito L, Heward J, Snook H, Chamberlain G, Rainbow DB, Hunter KM, Smith AN, Di Genova G, Herr MH, Dahlman I, Payne F, Smyth D, Lowe C, Twells RC, Howlett S, Healy B, Nutland S, Rance HE, Everett V, Smink LJ, Lam AC, Cordell HJ, Walker NM, Bordin C, Hulme J, Motzo C, Cucca F, Hess JF, Metzker ML, Rogers J, Gregory S, Allahabadia A, Nithiyananthan R, Tuomilehto-Wolf E, Tuomilehto J, Bingley P, Gillespie KM, Udlien DE, Ronningen KS, Guja C, Ionescu-Tirgoviste C, Savage DA, Maxwell AP, Carson DJ, Patterson CC, Franklyn JA, Clayton DG, Peterson LB, Wicker LS, Todd JA, Gough SC** 2003 Association of the T-cell regulatory gene CTLA4 with susceptibility to autoimmune disease. *Nature* 423:506–511
 152. **Vaidya B, Imrie H, Geatch DR, Perros P, Ball SG, Baylis PH, Carr D, Hurel SJ, James RA, Kelly WF, Kemp EH, Young ET, Weetman AP, Kendall-Taylor P, Pearce SH** 2000 Association analysis of the cytotoxic T lymphocyte antigen-4 (CTLA-4) and autoimmune reg-

- ulator-1 (AIRE-1) genes in sporadic autoimmune Addison's disease. *J Clin Endocrinol Metab* 85:688–691
153. Huang D, Liu L, Noren K, Xia SQ, Trifunovic J, Pirskanen R, Lefvert AK 1998 Genetic association of Ctl4 to myasthenia gravis with thymoma. *J Neuroimmunol* 88:192–198
 154. Donner H, Braun J, Seidl C, Rau H, Finke R, Ventz M, Walfish PG, Usadel KH, Badenhop K 1997 Codon 17 polymorphism of the cytotoxic T lymphocyte antigen 4 gene in Hashimoto's thyroiditis and Addison's disease. *J Clin Endocrinol Metab* 82:4130–4132
 155. Sale MM, Akamizu T, Howard TD, Yokota T, Nakao K, Mori T, Iwasaki H, Rich SS, Jennings-Gee JE, Yamada M, Bowden DW 1997 Association of autoimmune thyroid disease with a microsatellite marker for the thyrotropin receptor gene and CTLA-4 in a Japanese population. *Proc Assoc Am Physicians* 109:453–461
 156. Petrone A, Giorgi G, Mesturino CA, Capizzi M, Cascino I, Nistico L, Osborn J, Di Mario U, Buzzetti R 2001 Association of DRB1*04-DQB1*0301 haplotype and lack of association of two polymorphic sites at CTLA-4 gene with Hashimoto's thyroiditis in an Italian population. *Thyroid* 11:171–175
 157. Tomer Y, Greenberg DA, Barbesino G, Concepcion ES, Davies TF 2001 CTLA-4 and not CD28 is a susceptibility gene for thyroid autoantibody production. *J Clin Endocrinol Metab* 86:1687–1693
 158. Zaletel K, Krhin B, Gaberscek S, Pirnat E, Hojker S 2002 The influence of the exon 1 polymorphism of the cytotoxic T lymphocyte antigen 4 gene on thyroid antibody production in patients with newly diagnosed Graves' disease. *Thyroid* 12:373–376
 159. Vanderpump MPJ, Tunbridge WMG, French JM, Appleton D, Bates D, Clark F, Grimley Evans J, Hasan DM, Rodgers H, Tunbridge F, Young ET 1995 The incidence of thyroid disorders in the community: a twenty-year follow-up of the Whickham survey. *Clin Endocrinol (Oxf)* 43:55–68
 160. Kinjo Y, Takasu N, Komiyama I, Tomoyose T, Takara M, Kouki T, Shimajiri Y, Yabiku K, Yoshimura H 2002 Remission of Graves' hyperthyroidism and A/G polymorphism at position 49 in exon 1 of cytotoxic T lymphocyte-associated molecule-4 gene. *J Clin Endocrinol Metab* 87:2593–2596
 161. Yung E, Cheng PS, Fok TF, Wong GW 2002 CTLA-4 gene A-G polymorphism and childhood Graves' disease. *Clin Endocrinol (Oxf)* 56:649–653
 162. Allahabadia A, Heward JM, Nithiyananthan R, Gibson SM, Reuser TT, Dodson PM, Franklyn JA, Gough SC 2001 MHC class II region, CTLA4 gene, and ophthalmopathy in patients with Graves' disease. *Lancet* 358:984–985
 163. Vaidya B, Imrie H, Perros P, Dickinson J, McCarthy MI, Kendall-Taylor P, Pearce SH 1999 Cytotoxic T lymphocyte antigen-4 (CTLA-4) gene polymorphism confers susceptibility to thyroid associated orbitopathy. *Lancet* 354:743–744
 164. Marron MP, Zeidler A, Raffel LJ, Eckenrode SE, Yang JJ, Hopkins DI, Garchon HJ, Jacob CO, Serrano-Rios M, Martinez Larrad MT, Park Y, Bach JF, Rotter JI, Yang MC, She JX 2000 Genetic and physical mapping of a type 1 diabetes susceptibility gene (IDDM12) to a 100-kb phagemid artificial chromosome clone containing D2S72-CTLA4-D2S105 on chromosome 2q33. *Diabetes* 49:492–499
 165. Wood JP, Pani MA, Bieda K, Meyer G, Usadel KH, Badenhop K 2002 A recently described polymorphism in the CD28 gene on chromosome 2q33 is not associated with susceptibility to type 1 diabetes. *Eur J Immunogenet* 29:347–349
 166. Davies TF, Martin A, Concepcion ES, Graves P, Cohen L, Ben-Nun A 1991 Evidence of limited variability of antigen receptors on intrathyroidal T cells in autoimmune thyroid disease. *N Engl J Med* 325:238–244
 167. Demaine A, Welsh KI, Hawe BS, Farid NR 1987 Polymorphism of the T cell receptor β -chain in Graves' disease. *J Clin Endocrinol Metab* 65:643–646
 168. Weetman AP, So AK, Roe C, Walport MJ, Foroni L 1987 T-cell receptor α chain V region polymorphism linked to primary autoimmune hypothyroidism but not Graves' disease. *Hum Immunol* 20:167–173
 169. Uno H, Sasazuki H, Tamia H, Matsumoto H 1981 Two major genes, linked to HLA and Gm, control susceptibility to Graves' disease. *Nature* 292:768–770
 170. Nagataki S 1986 The interaction of MHC and Gm in liability to autoimmune thyroid disease. *Mol Biol Med* 3:73–84
 171. Nakao Y, Matsumoto H, Miyazaki T, Nishitani H, Takatsuki K, Kasukawa R, Nakayama S, Izumi S, Fujita T, Tsuji K 1980 IgG heavy chain allotypes (Gm) in atrophic and goitrous thyroiditis. *Clin Exp Immunol* 42:20–26
 172. Roman SH, Hubbard M, Rubinstein P, Failure to confirm standard HLA and Gm immunogenetic typing as a predictor of familial autoimmune thyroid disease. Program of the 71st Annual Meeting of The Endocrine Society, Seattle, WA, 1989 (Abstract)
 173. Fakhfakh F, Maalej A, Makni H, Abid M, Jouida J, Zouali M, Ayadi H 1999 Analysis of immunoglobulin VH and TCR β gene polymorphisms in a large family with thyroid autoimmune disorder. *Exp Clin Immunogenet* 16:185–191
 174. Tomer Y, Barbesino G, Keddache M, Greenberg DA, Davies TF 1997 Mapping of a major susceptibility locus for Graves' disease (GD-1) to chromosome 14q31. *J Clin Endocrinol Metab* 82:1645–1648
 175. Blakemore AIF, Watson PF, Weetman AP, Duff GW 1995 Association of Graves' disease with an allele of the interleukin-1 receptor antagonist gene. *J Clin Endocrinol Metab* 80:111–115
 176. Cuddihy RM, Bahn RS 1996 Lack of an association between alleles of interleukin-1 α and interleukin-1 receptor antagonist genes and Graves' disease in a North American Caucasian population. *J Clin Endocrinol Metab* 81:4476–4478
 177. Muhlberg T, Kirchberger M, Spitzweg C, Herrmann F, Heberling HJ, Heufelder AE 1998 Lack of association of Graves' disease with the A2 allele of the interleukin-1 receptor antagonist gene in a white European population. *Eur J Endocrinol* 138:686–690
 178. Heward J, Allahabadia A, Gordon C, Sheppard MC, Barnett AH, Franklyn JA, Gough SC 1999 The interleukin-1 receptor antagonist gene shows no allelic association with three autoimmune diseases. *Thyroid* 9:627–628
 179. Kamizono S, Hiromatsu Y, Seki N, Bednarczuk T, Matsumoto H, Kimura A, Itoh K 2000 A polymorphism of the 5' flanking region of tumour necrosis factor α gene is associated with thyroid-associated ophthalmopathy in Japanese. *Clin Endocrinol (Oxf)* 52:759–764
 180. Siegmund T, Usadel KH, Donner H, Braun J, Walfish PG, Badenhop K 1998 Interferon- γ gene microsatellite polymorphisms in patients with Graves' disease. *Thyroid* 8:1013–1017
 181. Rau H, Nicolay A, Usadel KH, Finke R, Donner H, Walfish PG, Badenhop K 1997 Polymorphisms of TAP1 and TAP2 genes in Graves' disease. *Tissue Antigens* 49:16–22
 182. Heward JM, Nithiyananthan R, Allahabadia A, Gibson S, Franklyn JA, Gough SC 2001 No association of an interleukin 4 gene promoter polymorphism with Graves' disease in the United Kingdom. *J Clin Endocrinol Metab* 86:3861–3863
 183. Ban Y, Taniyama M, Ban Y 2000 Vitamin D receptor gene polymorphism is associated with Graves' disease in the Japanese population. *J Clin Endocrinol Metab* 85:4639–4643
 184. Pani MA, Regulla K, Segni M, Hofmann S, Hufner M, Pasquino AM, Usadel KH, Badenhop K 2002 A polymorphism within the vitamin D-binding protein gene is associated with Graves' disease but not with Hashimoto's thyroiditis. *J Clin Endocrinol Metab* 87:2564–2567
 185. Tonacchera M, Pinchera A 2000 Thyrotropin receptor polymorphisms and thyroid diseases. *J Clin Endocrinol Metab* 85:2637–2639
 186. Cuddihy RM, Dutton CM, Bahn RS 1995 A polymorphism in the extracellular domain of the thyrotropin receptor is highly associated with autoimmune thyroid disease in females. *Thyroid* 5:89–95
 187. Kotsa KD, Watson PF, Weetman AP 1997 No association between a thyrotropin receptor gene polymorphism and Graves' disease in the female population. *Thyroid* 7:31–33
 188. Allahabadia A, Heward JM, Mijovic C, Carr-Smith J, Daykin J, Cockram C, Barnett AH, Sheppard MC, Franklyn JA, Gough SC 1998 Lack of association between polymorphism of the thyrotropin receptor gene and Graves' disease in United Kingdom and Hong Kong Chinese patients: case control and family-based studies. *Thyroid* 8:777–780
 189. Simanainen J, Kinch A, Westermark K, Winsa B, Bengtsson M, Schuppert F, Westermark B, Heldin NE 1999 Analysis of mutations in exon 1 of the human thyrotropin receptor gene: high frequency of the D36H and P52T polymorphic variants. *Thyroid* 9:7–11

190. **Kaczur V, Takacs M, Szalai C, Falus A, Nagy Z, Berencsi G, Balazs C** 2000 Analysis of the genetic variability of the 1st (CCC/ACC, P52T) and the 10th exons (bp 1012–1704) of the TSH receptor gene in Graves' disease. *Eur J Immunogenet* 27:17–23
191. **Chistyakov DA, Savost'yanov KV, Turakulov RI, Petunina NA, Trukhina LV, Kudinova AV, Balabolkin MI, Nosikov VV** 2000 Complex association analysis of Graves' disease using a set of polymorphic markers. *Mol Genet Metab* 70:214–218
192. **Rapoport B, Chazenbalk GD, Jaume JC, McLachlan SM** 1998 The thyrotropin (TSH) receptor: interaction with TSH and autoantibodies. *Endocr Rev* 19:673–716
193. **Tomer Y, Barbesino G, Greenberg DA, Concepcion ES, Davies TF** 1999 Mapping the major susceptibility loci for familial Graves' and Hashimoto's diseases: evidence for genetic heterogeneity and gene interactions. *J Clin Endocrinol Metab* 84:4656–4664
194. **De Roux N, Shields DC, Misrahi M, Ratanachaiyavong S, McGregor AM, Milgrom E** 1996 Analysis of the thyrotropin receptor as a candidate gene in familial Graves' disease. *J Clin Endocrinol Metab* 81:3483–3486
195. **Chistiakov DA, Savost'yanov KV, Turakulov RI, Petunina N, Balabolkin MI, Nosikov VV** 2002 Further studies of genetic susceptibility to Graves' disease in a Russian population. *Med Sci Monit* 8:CR180–CR184
196. **Muhlberg T, Herrmann K, Joba W, Kirchberger M, Heberling HJ, Heufelder AE** 2000 Lack of association of nonautoimmune hyperfunctioning thyroid disorders and a germline polymorphism of codon 727 of the human thyrotropin receptor in a European Caucasian population. *J Clin Endocrinol Metab* 85:2640–2643
197. **Ban Y, Greenberg DA, Concepcion ES, Tomer Y** 2002 A germline single nucleotide polymorphism at the intracellular domain of the human thyrotropin receptor does not have a major effect on the development of Graves' disease. *Thyroid* 12:1079–1083
198. **Pirro MT, De Filippis V, Di Cerbo A, Scillitani A, Liuzzi A, Tassi V** 1995 Thyroperoxidase microsatellite polymorphism in thyroid disease. *Thyroid* 5:461–464
199. **Tomer Y** 1997 Anti-thyroglobulin autoantibodies in autoimmune thyroid diseases: cross-reactive or pathogenic? *Clin Immunol Immunopathol* 82:3–11
200. **Roitt IM, Campbell PN, Doniach D** 1958 The nature of the thyroid autoantibodies present in patients with Hashimoto's thyroiditis (lymphadenoid goitre). *Biochem J* 69:248–254
201. **Ericsson UB, Christensen SB, Thorell J** 1985 A high prevalence of thyroglobulin autoantibodies in adults with and without thyroid disease as measured with a sensitive solid-phase immunosorbent radioassay. *Clin Immunol Immunopathol* 37:154–162
202. **Kuppers RC, Bresler HS, Lynne Burek C, Gleason SL, Rose NR** 1992 Immunodominant determinants of thyroglobulin associated with autoimmune thyroiditis. In: Bona CA, Kaushik AK, eds. *Molecular immunobiology of self-reactivity*. New York: Marcel Dekker; 247–284
203. **Vladutiu AO, Rose NR** 1971 Autoimmune murine thyroiditis: relation to histocompatibility (H-2) type. *Science* 174:1137–1139
204. **Beisel K, David CS, Giraldo AA, Kong Y-CM, Rose NR** 1982 Regulation of experimental autoimmune thyroiditis: mapping of susceptibility to the I-A subregion of the mouse H-2. *Immunogenetics* 15:427–430
205. **Kong YM, David CS, Lomo LC, Fuller BE, Motte RW, Giraldo AA** 1997 Role of mouse and human class II transgenes in susceptibility to and protection against mouse autoimmune thyroiditis. *Immunogenetics* 46:312–317
206. **Kong YC, McCormick DJ, Wan Q, Motte RW, Fuller BE, Giraldo AA, David CS** 1995 Primary hormonogenic sites as conserved autoepitopes on thyroglobulin in murine autoimmune thyroiditis. Secondary role of iodination. *J Immunol* 155:5847–5854
207. **Hutchings PR, Cooke A, Dawe K, Champion BR, Geysen M, Valerio R, Roitt IM** 1992 A thyroxine-containing peptide can induce murine experimental autoimmune thyroiditis. *J Exp Med* 175:869–872
208. **Wan Q, Motte RW, McCormick DJ, Fuller BE, Giraldo AA, David CS, Kong YM** 1997 Primary hormonogenic sites as conserved autoepitopes on thyroglobulin in murine autoimmune thyroiditis: role of MHC class II. *Clin Immunol Immunopathol* 85:187–194
209. **Texier B, Bedin C, Tang H, Camoin L, Laurent Winter C, Charreire J** 1992 Characterization and sequencing of a 40-amino-acid peptide from human thyroglobulin inducing experimental autoimmune thyroiditis. *J Immunol* 148:3405–3411
210. **Chronopoulou E, Carayanniotis G** 1992 Identification of a thyroiditogenic sequence within the thyroglobulin molecule. *J Immunol* 149:1039–1044
211. **Allen EM, Appel MC, Braverman LE** 1987 Iodine-induced thyroiditis and hypothyroidism in the hemithyroidectomized BB/W rat. *Endocrinology* 121:481–485
212. **Braley-Mullen H, Sharp GC, Medling B, Tang H** 1999 Spontaneous autoimmune thyroiditis in NOD.H-2 h4 mice. *J Autoimmun* 12:157–165
213. **Premawardhana LD, Lo SS, Phillips DI, Prentice LM, Rees-Smith B** 1994 Variability of serum thyroglobulin levels is determined by a major gene. *Clin Endocrinol (Oxf)* 41:725–729
214. **Tomer Y, Greenberg DA, Concepcion E, Ban Y, Davies TF** 2002 Thyroglobulin is a thyroid specific gene for the familial autoimmune thyroid diseases. *J Clin Endocrinol Metab* 87:404–407
215. **Grossman CJ, Roselle GA, Mendenhall CL** 1991 Sex steroid regulation of autoimmunity. *J Steroid Biochem Mol Biol* 40:649–659
216. **Ando T, Imaizumi M, Graves PN, Unger P, Davies TF** 2002 Intrathyroidal fetal microchimerism in Graves' disease. *J Clin Endocrinol Metab* 87:3315–3320
217. **Paavonen T** 1994 Hormonal regulation of immune responses. *Ann Med* 26:255–258
218. **Ansar AS, Penhale WJ, Talal N** 1985 Sex hormones, immune responses, and autoimmune diseases. Mechanisms of sex hormone action. *Am J Pathol* 121:531–551
219. **Okayasu I, Kong YM, Rose NR** 1981 Effect of castration and sex hormones on experimental autoimmune thyroiditis. *Clin Immunol Immunopathol* 20:240–245
220. **Kanda N, Tamaki K** 1999 Estrogen enhances immunoglobulin production by human PBMCs. *J Allergy Clin Immunol* 103:282–288
221. **Ansar AS, Young PR, Penhale WJ** 1983 The effects of female sex steroids on the development of autoimmune thyroiditis in thymectomized and irradiated rats. *Clin Exp Immunol* 54:351–358
222. **Kim S, Liva SM, Dalal MA, Verity MA, Voskuhl RR** 1999 Estriol ameliorates autoimmune demyelinating disease: implications for multiple sclerosis. *Neurology* 52:1230–1238
223. **de Kerdanet M, Lucas J, Lemeze F, Lecornu M** 1994 Turner's syndrome with X-isochromosome and Hashimoto's thyroiditis. *Clin Endocrinol (Oxf)* 41:673–676
224. **Ivarsson SA, Ericsson UB, Nilsson KO, Gustafsson J, Hagenas L, Hager A, Moell C, Tuvemo T, Westphal O, Albertsson-Wikland K** 1995 Thyroid autoantibodies, Turner's syndrome and growth hormone therapy. *Acta Paediatr* 84:63–65
225. **Medeiros CC, Marini SH, Baptista MT, Guerra Jr G, Maciel-Guerra AT** 2000 Turner's syndrome and thyroid disease: a transverse study of pediatric patients in Brazil. *J Pediatr Endocrinol Metab* 13:357–362
226. **Elsheikh M, Wass JA, Conway GS** 2001 Autoimmune thyroid syndrome in women with Turner's syndrome—the association with karyotype. *Clin Endocrinol (Oxf)* 55:223–226
227. **Barbesino G, Tomer Y, Concepcion ES, Davies TF, Greenberg D** 1998 Linkage analysis of candidate genes in autoimmune thyroid disease. 2. Selected gender-related genes and the X-chromosome. *J Clin Endocrinol Metab* 83:3290–3295
228. **Tomer Y, Ban Y, Concepcion E, Barbesino G, Villanueva R, Greenberg DA, Davies TF** Common and unique susceptibility loci in Graves' and Hashimoto's diseases: results of whole genome screening in a dataset of 102 multiplex multi-generational families. *Am J Hum Genet*, in press
229. **Imrie H, Vaidya B, Perros P, Kelly WF, Toft AD, Young ET, Kendall-Taylor P, Pearce SHS** 2001 Evidence for a Graves' disease susceptibility locus at chromosome Xp11 in a United Kingdom population. *J Clin Endocrinol Metab* 86:626–630
230. **Stewart JJ** 1998 The female X-inactivation mosaic in systemic lupus erythematosus. *Immunol Today* 19:352–357
231. **Chitnis S, Monteiro J, Glass D, Apatoff B, Salmon J, Concannon P, Gregersen PK** 2000 The role of X-chromosome inactivation in female predisposition to autoimmunity. *Arthritis Res* 2:399–406
232. **Weetman AP, Jenkins RC** 2002 Disease associations with autoimmune thyroid disease. *Thyroid* 12:977–988

233. **Gray RS, Elton RA, Clarke BF** 1983 Familial distribution of thyroid disease and diabetes: further evidence for aetiological heterogeneity of diabetes mellitus. *Q J Med* 52:244–255
234. **Payami H, Joe S, Thomson G** 1989 Autoimmune thyroid disease in type 1 diabetes. *Genet Epidemiol* 6:137–141
235. **Torfs CP, King M-C, Bing H, Malmgren J, Grumet FC** 1986 Genetic interrelationship between insulin-dependent diabetes mellitus, the autoimmune thyroid diseases, and rheumatoid arthritis. *Am J Hum Genet* 38:170–187
236. **Tomer Y, Greenberg DA, Davies TF** 1999 The genetic susceptibility to type 1 (insulin dependent) diabetes mellitus and autoimmune thyroid diseases: from epidemiological observations to gene mapping. In: Volpe R, ed. *Contemporary endocrinology: autoimmune endocrinopathies*. Totowa, NJ: Humana Press; 57–90
237. **Kordonouri O, Klinghammer A, Lang EB, Gruters-Kieslich A, Grabert M, Holl RW** 2002 Thyroid autoimmunity in children and adolescents with type 1 diabetes: a multicenter survey. *Diabetes Care* 25:1346–1350
238. **Kordonouri O, Deiss D, Danne T, Dorow A, Bassir C, Gruters-Kieslich A** 2002 Predictivity of thyroid autoantibodies for the development of thyroid disorders in children and adolescents with type 1 diabetes. *Diabet Med* 19:518–521
239. **Alvarez-Marfany M, Roman SH, Drexler AJ, Robertson C, Stagnaro-Green A** 1994 Long-term prospective study of postpartum thyroid dysfunction in women with insulin dependent diabetes mellitus. *J Clin Endocrinol Metab* 79:10–16
240. **Bech K, Hoier-Madsen M, Feldt-Rasmussen U, Jensen BM, Molsted-Pedersen L, Kuhl C** 1991 Thyroid function and autoimmune manifestations in insulin-dependent diabetes mellitus during and after pregnancy. *Acta Endocrinol (Copenh)* 124:534–539
241. **Weetman AP** 1994 Insulin-dependent diabetes mellitus and postpartum thyroiditis: an important association. *J Clin Endocrinol Metab* 79:7–9
242. **McCanlies E, O'Leary LA, Foley TP, Kramer MK, Burke JP, Libman A, Swan JS, Steenkiste AR, McCarthy BJ, Trucco M, Dorman JS** 1998 Hashimoto's thyroiditis and insulin-dependent diabetes mellitus: differences among individuals with and without abnormal thyroid function. *J Clin Endocrinol Metab* 83:1548–1551
243. **Cox NJ, Wapelhorst B, Morrison VA, Johnson L, Pinchuk L, Spielman RS, Todd JA, Concannon P** 2001 Seven regions of the genome show evidence of linkage to type 1 diabetes in a consensus analysis of 767 multiplex families. *Am J Hum Genet* 69:820–830
244. **Luo DF, Bui MM, Muir A, Maclaren NK, Thomson G, She JX** 1995 Affected sibpair mapping of a novel susceptibility gene to insulin-dependent diabetes mellitus (IDDM8) on chromosome 6q25–27. *Am J Hum Genet* 57:911–919
245. **Luo DF, Buzzetti R, Rotter JL, Maclaren NK, Raffel LJ, Nistico L, Giovannini C, Pozzilli P, Thomson G, She JX** 1996 Confirmation of three susceptibility genes to insulin-dependent diabetes mellitus: IDDM4, IDDM5 and IDDM8. *Hum Mol Genet* 5:693–698
246. **Allahabadia A, Heward J, Carr-Smith J, Daykin J, Barnett AH, Sheppard MC, Franklyn JA, Gough SC** 1999 Sharing of susceptibility loci between autoimmune diseases: lack of association of the insulin gene region with Graves' disease. *Thyroid* 9:317–318
247. **Vaidya B, Imrie H, Perros P, Young ET, Kelly WF, Carr D, Large DM, Toft AD, Kendall-Taylor P, Pearce SH** 2000 Evidence for a new Graves disease susceptibility locus at chromosome 18q21. *Am J Hum Genet* 66:1710–1714
248. **Tomer Y, Barbesino G, Greenberg DA, Concepcion ES, Davies TF** 1998 Linkage analysis of candidate genes in autoimmune thyroid disease. 3. Detailed analysis of chromosome 14 localizes GD-1 close to MNG-1. *J Clin Endocrinol Metab* 83:4321–4327
249. **Biunno I, Appierto V, Cattaneo M, Leone BE, Balzano GP, Succi C, Saccone S, Letizia A, Della Valle G, Sgaramella V** 1997 Isolation of a pancreas-specific gene located on human chromosome 14q31: expression analysis in human pancreatic ductal carcinomas. *Genomics* 46:284–286
250. **Ban Y, Taniyama M, Tozaki T, Yanagawa T, Tomita M, Ban Y** 2001 SEL1L microsatellite polymorphism in Japanese patients with autoimmune thyroid diseases. *Thyroid* 11:335–338
251. **Pearce SH, Vaidya B, Imrie H, Perros P, Kelly WF, Toft AD, McCarthy MI, Young ET, Kendall-Taylor P** 1999 Further evidence for a susceptibility locus on chromosome 20q13.11 in families with dominant transmission of Graves disease. *Am J Hum Genet* 65:1462–1465
252. **Tomer Y, Concepcion E, Greenberg DA** 2002 A C/T single nucleotide polymorphism in the region of the CD40 gene is associated with Graves' disease. *Thyroid* 12:1129–1135
253. **Durie FH, Foy TM, Masters SR, Laman JD, Noelle RJ** 1994 The role of CD40 in the regulation of humoral and cell-mediated immunity. *Immunol Today* 15:406–411
254. **Banchereau J, Bazan F, Blanchard D, Briere F, Galizzi JP, van Kooten C, Liu YJ, Rousset F, Saeland S** 1994 The CD40 antigen and its ligand. *Annu Rev Immunol* 12:881–922
255. **Foy TM, Aruffo A, Bajorath J, Buhlmann JE, Noelle RJ** 1996 Immune regulation by CD40 and its ligand GP39. *Annu Rev Immunol* 14:591–617
256. **Carayanniotis G, Masters SR, Noelle RJ** 1997 Suppression of murine thyroiditis via blockade of the CD40-CD40L interaction. *Immunology* 90:421–426
257. **Villanueva R, Tomer Y, Greenberg DA, Mao C, Concepcion ES, Tucci S, Estilo G, Davies TF** 2002 Autoimmune thyroid disease susceptibility loci in a large Chinese family. *Clin Endocrinol (Oxf)* 56:45–51
258. **Eng C** 1996 Seminars in medicine of the Beth Israel Hospital, Boston. The RET proto-oncogene in multiple endocrine neoplasia type 2 and Hirschsprung's disease. *N Engl J Med* 335:943–951
259. **Puxeddu E, Fagin JA** 2001 Genetic markers in thyroid neoplasia. *Endocrinol Metab Clin North Am* 30:493–513
260. **Aitman TJ, Todd JA** 1995 Molecular genetics of diabetes mellitus. *Baillière's Clin Endocrinol Metab* 9:631–656
261. **Kouki T, Sawai Y, Gardine CA, Fisfalen ME, Alegre ML, DeGroot LJ** 2000 CTLA-4 gene polymorphism at position 49 in exon 1 reduces the inhibitory function of CTLA-4 and contributes to the pathogenesis of Graves' disease. *J Immunol* 165:6606–6611
262. **Xu Y, Graves P, Tomer Y, Davies T** 2002 CTLA-4 and autoimmune thyroid disease: lack of influence of the A49G signal peptide polymorphism on functional recombinant human CTLA-4. *Cell Immunol* 215:133–140
263. **Buus S, Sette A, Grey HM** 1987 The interaction between protein-derived immunogenic peptides and Ia. *Immunol Rev* 98:115–141
264. **Faas S, Trucco M** 1994 The genes influencing the susceptibility to IDDM in humans. *J Endocrinol Invest* 17:477–495
265. **Morel PA, Dorman JS, Todd JA, McDevitt HO, Trucco M** 1988 Aspartic acid at position 57 of the HLA-DQ β -chain protects against type I diabetes: a family study. *Proc Natl Acad Sci USA* 85:8111–8115
266. **Brown JH, Jardetzky T, Gorga JC, Stern LJ, Urban RG, Strominger JL, Wiley DC** 1993 Three-dimensional structure of the human class II histocompatibility antigen HLA-DR1. *Nature* 364:33–39
267. **Lee KH, Wucherpfennig KW, Wiley DC** 2001 Structure of a human insulin peptide-HLA-DQ8 complex and susceptibility to type 1 diabetes. *Nat Immunol* 2:501–507
268. **Wucherpfennig KW** 2001 Insights into autoimmunity gained from structural analysis of MHC-peptide complexes. *Curr Opin Immunol* 13:650–656
269. **Sawai Y, DeGroot LJ** 2000 Binding of human thyrotropin receptor peptides to a Graves' disease-predisposing human leukocyte antigen class II molecule. *J Clin Endocrinol Metab* 85:1176–1179
270. **Hanafusa T, Pujol Borrell R, Chiovato L, Russell RC, Doniach D, Bottazzo GF** 1983 Aberrant expression of HLA-DR antigen on thyrocytes in Graves' disease: relevance for autoimmunity. *Lancet* 2:1111–1115
271. **Bottazzo GF, Pujol Borrell R, Hanafusa T, Feldmann M** 1983 Role of aberrant HLA-DR expression and antigen presentation in induction of endocrine autoimmunity. *Lancet* 2:1115–1119
272. **Davies TF** 1985 Cocultures of human thyroid monolayer cells and autologous T cells: impact of HLA class II antigen expression. *J Clin Endocrinol Metab* 61:418–422
273. **Londei M, Lamb JR, Bottazzo GF, Feldmann M** 1984 Epithelial cells expressing aberrant MHC class II determinants can present antigen to cloned human T cells. *Nature* 312:639–641
274. **Tomer Y, Davies TF** 2000 The genetics of familial and non-familial hyperthyroid Graves' disease. In: Rapoport B, McLachlan SM, eds. *Graves' disease: pathogenesis and treatment*. Boston: Kluwer Academic Publishers; 19–41

275. **Davies TF, Piccinini LA** 1987 Intrathyroidal MHC class II antigen expression and thyroid autoimmunity. *Endocrinol Metab Clin North Am* 16:247–268
276. **Neufeld DS, Platzer M, Davies TF** 1989 Reovirus induction of MHC class II antigen in rat thyroid cells. *Endocrinology* 124: 543–545
277. **Belfiore A, Mauerhoff T, Pujol Borrell R, Badenhoop K, Buscema M, Mirakian R, Bottazzo GF** 1991 *De novo* HLA class II and enhanced HLA class I molecule expression in SV40 transfected human thyroid epithelial cells. *J Autoimmun* 4:397–414
278. **Shimojo N, Kohno Y, Yamaguchi K, Kikuoka S, Hoshioka A, Niimi H, Hirai A, Tamura Y, Saito Y, Kohn LD, Tahara K** 1996 Induction of Graves-like disease in mice by immunization with fibroblasts transfected with the thyrotropin receptor and a class II molecule. *Proc Natl Acad Sci USA* 93:11074–11079
279. **Kita M, Ahmad L, Marians RC, Vlase H, Unger P, Graves PN, Davies TF** 1999 Regulation and transfer of a murine model of thyrotropin receptor antibody mediated Graves' disease. *Endocrinology* 140:1392–1398
280. **Davies TF, Bermas B, Platzer M, Roman SH** 1985 T-cell sensitization to autologous thyroid cells and normal non-specific suppressor T-cell function in Graves' disease. *Clin-Endocrinol (Oxf)* 22:155–167
281. **Eguchi K, Otsubo T, Kawabe K, Ueki Y, Fukuda T, Mayumi M, Shimomura C, Ishikawa N, Nakao H, Ito K, Morimoto C, Nagataki S** 1987 The remarkable proliferation of helper T cell subset in response to autologous thyrocytes and intrathyroidal T cells from patients with Graves' disease. *Isr J Med Sci* 70:403–410
282. **Migita K, Eguchi K, Otsubo T, Kawakami A, Nakao H, Ueki Y, Shimomura C, Kurata A, Fukuda T, Matsunaga M, Ishikawa N, Ito K, Nagataki S** 1990 Cytokine regulation of HLA on thyroid epithelial cells. *Clin Exp Immunol* 82:548–552
283. **Weetman AP, McGregor AM** 1994 Autoimmune thyroid disease: further developments in our understanding. *Endocr Rev* 15: 788–830
284. **Huang D, Giscombe R, Zhou Y, Pirskanen R, Lefvert AK** 2000 Dinucleotide repeat expansion in the CTLA-4 gene leads to T cell hyper-reactivity via the CD28 pathway in myasthenia gravis. *J Neuroimmunol* 105:69–77
285. **Holopainen PM, Partanen J** 2001 Technical note: linkage disequilibrium and disease-associated CTLA-4 gene polymorphisms. *J Immunol* 167:2457–2458
286. **Bagchi N, Brown TR, Urduvia E, Sundick RS** 1985 Induction of autoimmune thyroiditis in chickens by dietary iodine. *Science* 230: 325–327
287. **Kahaly GJ, Dienes HP, Beyer J, Hommel G** 1998 Iodide induces thyroid autoimmunity in patients with endemic goitre: a randomised, double-blind, placebo-controlled trial. *Eur J Endocrinol* 139: 290–297
288. **Papanastasiou L, Alevizaki M, Pipingos G, Mantzos E, Tseloni-Balafouta S, Koutras DA** 2000 The effect of iodine administration on the development of thyroid autoimmunity in patients with nontoxic goiter. *Thyroid* 10:493–497
289. **dos Remedios LV, Weber PM, Feldman R, Schurr DA, Tsoi TG** 1980 Detecting unsuspected thyroid dysfunction by the free thyroxine index. *Arch Intern Med* 140:1045–1049
290. **Stenszky V, Kozma L, Balazs C, Rochkitz S, Bear JC, Farid NR** 1985 The genetics of Graves' disease: HLA and disease susceptibility. *J Clin Endocrinol Metab* 61:735–740
291. **Freckler M, Mercer G, Skanes VM, Farid NR** 1988 Major histocompatibility complex (MHC) factors predisposing to and protecting against Graves' eye disease. *Autoimmunity* 1:307–315
292. **McGregor A, Rees Smith B, Hall R, Petersen M, Miller M, Dewar P** 1980 Prediction of relapse in hyperthyroid Graves' disease. *Lancet* i:1101–1103
293. **Weetman AP, So AK, Warner CA, Foroni L, Fells P, Shine B** 1988 Immunogenetics of Graves' ophthalmopathy. *Clin Endocrinol (Oxf)* 28:619–628
294. **Chen QY, Huang W, She JX, Baxter F, Volpe R, Maclaren NK** 1999 HLA-DRB1*08, DRB1*03/DRB3*0101, and DRB3*0202 are susceptibility genes for Graves' disease in North American Caucasians, whereas DRB1*07 is protective. *J Clin Endocrinol Metab* 84:3182–3186
295. **Hawkins BR, Ma JT, Lam KS, Wang CC, Yeung RT** 1985 Association of HLA antigens with thyrotoxic Graves' disease and periodic paralysis in Hong Kong Chinese. *Clin Endocrinol (Oxf)* 23: 245–252
296. **Wong GWK, Cheng SH, Dorman JS** 1999 The HLA-DQ associations with Graves' disease in Chinese children. *Clin Endocrinol (Oxf)* 50:493–495
297. **Sasazuki T, Nishimura Y, Muto M, Ohta N** 1983 HLA-linked genes controlling immune response and disease susceptibility. *Immunol Rev* 70:51–75
298. **Dong RP, Kimura A, Okubo R, Shinagawa H, Tamai H, Nishimura Y, Sasazuki T** 1992 HLA-A and DPB1 loci confer susceptibility to Graves' disease. *Hum Immunol* 35:165–172
299. **Cho BY, Rhee BD, Lee DS, Lee MS, Kim GY, Lee HK, Koh CS, Min HK, Lee M** 1987 HLA and Graves' disease in Koreans. *Tissue Antigens* 30:119–121
300. **Tandon N, Mehra NK, Taneja V, Vaidya MC, Kochupillai N** 1990 HLA antigens in Asian Indian patients with Graves' disease. *Clin Endocrinol (Oxf)* 33:21–26
301. **Sridama V, Hara Y, Fauchet R, DeGroot LJ** 1987 HLA immunogenetic heterogeneity in Black American patients with Graves' disease. *Arch Intern Med* 147:229–231
302. **Chen QY, Nadell D, Zhang XY, Kukreja A, Huang YJ, Wise J, Svec F, Richards R, Friday KE, Vargas A, Gomez R, Chalew S, Lan MS, Tomer Y, Maclaren NK** 2000 The human leukocyte antigen HLA DRB3*020/DQA1*0501 haplotype is associated with Graves' disease in African Americans. *J Clin Endocrinol Metab* 85:1545–1549
303. **Omar MA, Hammond MG, Desai RK, Motala AA, Aboo N, Seedat MA** 1990 HLA class I and II antigens in South African blacks with Graves' disease. *Clin Immunol Immunopathol* 54:98–102
304. **Allen EM, Hsueh WC, Sabra MM, Pollin TI, Ladenson PW, Silver KD, Mitchell BD, Shuldiner AR** 2003 A genome-wide scan for autoimmune thyroiditis in the Old Order Amish: replication of genetic linkage on chromosome 5q11.2-q14.3 *J Clin Endocrinol Metab* 88:1292–1296
305. **Bogner U, Badenhoop K, Peters H, Schmiege D, Mayr WR, Usadel KH, Schleusener H** 1992 HLA-DR/DQ gene variation in non-goitrous autoimmune thyroiditis at the serological and molecular level. *Autoimmunity* 14:155–158
306. **Ito M, Tanimoto M, Kamura H, Yoneda M, Morishima Y, Yamachi K, Itatsu T, Takatsuki K, Saito H** 1989 Association of HLA antigen and restriction fragment length polymorphism of T cell receptor β -chain gene with Graves' disease and Hashimoto's thyroiditis. *J Clin Endocrinol Metab* 69:100–104
307. **Alkhateeb A, Stetler GL, Old W, Talbert J, Uhlhorn C, Taylor M, Fox A, Miller C, Dills DG, Ridgway EC, Bennett DC, Fain PR, Spritz RA** 2002 Mapping of an autoimmunity susceptibility locus (AIS1) to chromosome 1p31.3-p32.2. *Hum Mol Genet* 11:661–667
308. **Tomer Y, Davies TF** 2002 Genetic factors relating to the thyroid with emphasis on complex diseases. In: Waas JAH, Shalet SM, eds. *Oxford textbook of endocrinology and diabetes*. Oxford, UK: Oxford University Press; 358